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Oxidation of several triterpenic diene and triene systems. Oxidative cleavage to obtain chiral intermediates for drimane and phenanthrene semi-synthesis

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Abstract—An exhaustive oxidation study has been made with ozone, MCPBA and/or $NaIO_4/RuCl_3$ of several triterpenic diene and triene compounds from oleanolic and maslinic acids obtained from olive-oil pressing. Through several oxidative cleavages of the opened C-ring of these oleantrienes, different significant decalin-type chiral synthons were achieved. These sesquiterpene and *nor*-sesquiterpene products are of great interest because by means of several simple reactions they could lead to drimane, phenanthrene and tricyclic triterpene compounds. © 2004 Elsevier Ltd. All rights reserved.

1. Introduction

Triterpenes are a large family of pentacyclic compounds obtained biosynthetically by cyclic reactions from squalene.¹ Oleanolic (3 β -hydroxy-12-oleanen-28-oic acid)² and maslinic (2 α ,3 β -dihydroxy-12-oleanen-28-oic acid)² acids belong to these kinds of natural products widely found in nature, which could be useful to semi-synthesize other biologically or chemically significant compounds.³ The presence of these two oleanene acids in olive-pressing residues has been frequently reported. A method to obtain large amounts of both compounds from these solid wastes has been reported by our group.⁴

Due to their remarkable biological activities and their olfactory and fixative properties, drimane sesquiterpenes or related compounds are being vigorously sought by the scientific community.^{5,6} Likewise, some intermediates useful in the synthesis of drimane sesquiterpenes, such as copalol or albicanol, have been produced.⁷ These kinds of products can be obtained by degrading of different terpenes such as sclareol, some abietic acids and glycyrrhetinic acid,⁸ which are rarer and more expensive than oleanolic and maslinic acids.

Previous works have dealt with reactivity and rearrangement of several derivatives of oleanolic and maslinic acids, reactions which have provided high yields of several interesting A-ring contracted products.⁹ In addition, our group reported the isolation and unequivocal structural assignments of a pair of sulphur diastereomeric cyclic sulphites between C-2 and C-3 of maslinic acid.¹⁰ An interesting synthon for the A- and B-rings of the 4-azasteroids was also semi-synthesized. Recently, we have reported the formation of several C-ring derivatives and the cleavage of the triterpenic molecule.^{11,12}

In the present paper, we carry out an oxidative study of above-mentioned derivatives in C-ring with the final aim of cleaving the triterpenic molecule and to produce interesting chiral synthons. In this sense, dienes 3 and 4 were ozonized, triggering epoxidations, hydroxylations and lactonizations in one or the two double bonds of the C-ring. Moreover, the treatment of cis-triene 5 with some oxidative reagents such as NaIO₄/RuCl₃, MCPBA and ozone led to the same type of derivatives. In turn, oxidation of *trans*-triene 11 with NaIO₄/ RuCl₃ gave rise to the obtention of two sesquiterpene fragments (45 and 46) by C-ring cleavage of the triterpene molecule. Similarly, epoxidation of exocyclic triene 16 and subsequent ozonization of the major product yielded two nor-sesquiterpene compounds (53 and 54) and two C-16 fragments (55 and 56). The products formed from A- and B-rings of the original molecule (compounds 45, 53 and 54) could be used as a substrate for the semi-synthesis of significant drimane compounds, such as warburganal, polygodial, etc., since it has structure of 3B-hydroxydrimenal.⁸ Also, the fragments derived from D- and E-rings (products 46, 55 and 56) could be transformed into several phenanthrenes, which act as estrogen receptor modulators for the treatment or prevention of a variety of conditions related to estrogen functioning, including bone loss,

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Figure 1. Structures of products 1-16.

osteoporosis, cardiovascular disease, some types of cancer, etc.¹³ In addition, these fragments could be used as starting material in the semi-synthesis of natural tricyclic triterpenoids Achilleol B¹⁴ and Camelliol C.¹⁵

2. Results and discussion

Oleanolic and maslinic acids were isolated from solid wastes resulting from olive-oil production. Dienes 1-4 were obtained from the corresponding oleanolic and maslinic esters by a key bromination/dehydrobromination process with NBS (Fig. 1). These products were converted into trienes 5-8 by a photochemical reaction with a high-pressure Hg street lamp in a borosilicate flask, whereas trienes 9-12 were formed starting from trienes 5-8 by chemical isomerization with iodine, and trienes 13-16 by photochemical isomerization in a quartz flask. All these processes were described in previous reports.^{11,12}

Ozonolysis of diene **3** at a low temperature (-78 °C) for 15 min gave rise to several oxidized products (Scheme 1).

The major product, compound 17, was a lactone formed by the attack of the carboxymethyl group at C-28 on C-13 from the β face and opening of the α -epoxide previously formed between C-12 and C-13 by the ozonolysis process. The C-9/ C-11 double bond remained unaltered. This reaction was previously observed in the ozonization¹² and photochemical lactonization¹⁶ of oleanolic acid. In turn, product 18 was produced by epoxidation of the double bonds of the starting material both from the α face, which was the most accessible one. Meanwhile, compound 19 presented a taraxerene structure formed from product 18 by a concerted mechanism. It took place the opening of the epoxide between C-12/C-13, the migration of the methyl group at C-14 to C-13 by the α -face and the formation of a double bond between C-14 and C-15. This way, a β -hydroxyl group was situated at C-12, whereas the α -epoxide between C-9 and C-11 remained unaltered. This change of skeleton was supported by the spectroscopic properties of product 19 and was also observed in the reaction of methyl oleanate with NBS.¹² Finally, compound **20** was formed by the same mechanism as lactone 17 was, but with the epoxidation of the C-9/C-11 double bond also occurring. The structure of



Scheme 1. Reagents and conditions: (a) O₃/CH₂Cl₂/-78 °C/15 min/Me₂S/3 h 17 (30%), 18 (20%), 19 (15%) and 20 (7%).

these products suggested that for both double bonds the less hindered face would be the α face.

Likewise, the ozonolysis of diene **4** in MeOH/CH₂Cl₂ 4:1 at -78 °C led to several oxidized products, which were similar to those obtained by the ozonization of diene **3** (Scheme 2). Moreover, the major product, lactone **21**, was the equivalent compound to lactone **17**. Oxirane **22** was formed by epoxidation of the double bond between C-12 and C-13 from the less hindered face and the α , β -unsaturated ketone **23** was the result of the evolution of epoxide **22**. According to these structures, we conclude that the C-12/C-13 double bond would be more accessible by the reagent than the C-9/C-11 double bond in the oxidative process. Furthermore, the stereochemistry of all the epoxide and hydroxyl groups of products **17–22** was established from their spectroscopic characteristics.

In an attempt to cleave the triterpenic molecule by the C-ring, triene **5**, obtained from diene **1** by a photochemical reaction, 11,12 was treated with ozone under different conditions. In all cases, complex mixtures of products, very difficult to separate chromatographically, resulted. When pyridine was used as the co-solvent, only compound **24** was identified (Scheme 3, path a). This product was



Scheme 2. Reagents and conditions: (a) O₃/CH₂Cl₂:MeOH/-78 °C/ 10 min/thiodipropionic acid/10 min **21** (40%), **22** (20%) and **23** (12%).

formed by epoxidation of the most substituted double bonds of the molecule and hydrolysis of the oxirane group between C-13 and C-14. Subsequent oxidation of the corresponding hydroxyl groups at these positions gave rise to the cleavage of C-13/C-14 double bond, rendering the diketone **24**. Moreover, triene **5** was treated with NaIO₄/RuCl₃ at room temperature, giving good yield (80%) in cyclic ether **25** (Scheme 3, path b). The formation of this product could be based on the epoxidation of the C-8/C-9 and C-13/C-14 double bonds of the molecule and the hydrolysis of the epoxide group between C-13 and C-14. The attack of the hydroxyl group at C-13 on C-9 from the β face and opening of the C-8 α /C-9 α oxirane group yielded product **25**.

The presence of the hydroxyl groups and the cyclic ether in 25 was consistent with the spectroscopic properties of this product. Finally, triene 5 was epoxidized with MCPBA, rendering a mixture of several oxidized products with different yields depending on the reaction conditions (Scheme 3, path c) (Table 1). At low temperature (-78 °C), only product 26 resulted (25%), which had been epoxidized in C-8/C-9 from the α face. When the temperature rose to -40 °C, epoxide 26 was isolated in a good yield (70%). At -20 °C, in addition to compound **26**, whose yield had decreased, products 27, 28 and 29 were obtained. Compounds 27 and 28 had an epoxide group by the α face in C-8/C-9; however, while 27 had a new oxirane group also by the α face in C-13/C-14, in product 28 this group had the opposite configuration in this position. Product 29 was epoxidized in C-8/C-9 from the α face (as in the latter three compounds) and a lactone system was formed between C-28 and C-13 by the same mechanism as for products 17, 20 and 21. When the reaction was maintained at 0 °C for 1 h, two more epoxides were isolated. Product **30** had been epoxidized in C-8/C-9 from the β face, whereas the oxirane group in C-13/C-14 had the opposite configuration. In turn, product 31 has an epoxide group between C-8/C-9 by the α face and two hydroxyl groups at C-13 and C-14. Finally, at room temperature the epoxidation reaction rendered the same products as at 0 °C, but products 26 and 27 were obtained in a slightly better yield.

Moreover, when the reaction with MCPBA was carried out over monoepoxide 26 at -40 °C the major product was 27 and compounds 28 and 31 were also obtained (Scheme 4). At room temperature, the epoxidation of 26 gave rise to the same products as the treatment of triene 5 with MCPBA but with different yields, in addition to two new oxidized products 32 and 33 (Table 2). These compounds had similar structure as cyclic ether 25, but in 32 the hydroxyl group at C-8 had dehydrated with a H-7 hydrogen and in 33, the new double bond was formed between C-8 and C-26. As could be observed, product 30 was not isolated from the epoxidation of monoepoxide 26. This supported the β configuration attributed to the oxirane group between C-8 and C-9. As a mean to protect the hydroxyl group on C-3 of monoepoxide 26, an acetylation process was carried out with Ac_2O/Py , rendering acetate 34. This compound was also epoxidized, giving rise to the corresponding acetylated derivatives of products 27, 28, 29 and 31, compounds 35, 36, 37 and 38, respectively. In order to cleave the C-ring of the triterpenic molecule, products 35 and 37 were submitted to several oxidative reactions, yielding complex mixtures of



Scheme 3. Reagents and conditions: (a) $O_3/CH_2Cl_2/Py 2:1/-78$ °C/30 min 24 (12%); (b) $NaIO_4/RuCl_3/CCl_4/CH_3CN/H_2O/room$ temperature/8 h 25 (80%); (c) $MCPBA/CH_2Cl_2$ 26, 27, 28, 29, 30 and 31 (Table 1).

polar compounds very difficult to separate chromatographically.

With the same aim, ozonolysis of monoepoxide 26 was achieved (Scheme 5). When this reaction took place in CH₂Cl₂, only product **31** was obtained in an acceptable yield (60%). As this compound appeared to be suitable for the cleavage of the C-ring, it was ozonized again, rendering ketone **39**. In turn, when the ozonolysis of monoepoxide **26** was carried out in CH₂Cl₂/Py 10:1, ketone 40 was isolated with good yield (80%). The subsequent treatment of this product with ozone did not gave the expected results, since this reaction led to product 39 and lactone 41. These compounds resulted from the attack of the carboxymethyl group at C-28 on C-13 from the β face and the opening of the α epoxide between C-13 and C-14. In addition, a complex mixture of polar compounds was obtained. It was treated with diazomethane in order to form the corresponding methyl esters, but this reagent did not simplify the mixture.

Table 1. Yields (%) of products obtained by epoxidation of triene 5 at different temperatures (°C) and times (h)

Т	Time	26	27	28	29	30	31
-78	24	25	_	_	_	_	_
-40	12	70	_	_	_	_	_
-20	8	40	25	5	5		_
0	1	30	30	10	5	5	5
Room temperature	0.5	35	35	10	5	5	5

In an effort to avoid the participation of hydroxyl group at C-3 in the ozonolysis process, acetylated derivative **34** was ozonized. This reaction rendered the already isolated products **37** (acetyl derivative of **29**) and **38** (acetylated derivative of **31**) with different yields depending on reaction conditions. Table 3 shows that at -78 °C the major product



Scheme 4. Reagents and conditions: (a) MCPBA/CH₂Cl₂ 27, 28, 29, 31, 32 and 33 (Table 2).

 Table 2. Yields (%) of products obtained by epoxidation of monoepoxide

 26 at different temperatures (°C) and times (h)

Т	Time	27	28	29	31	32	33
-40 Room temperature	12 1	75 15	15 10	<u> </u>	7 30	5	5

In addition, since oxidation reactions carried out using several compounds with the central double bond in *Z*-disposition did not allow the cleavage of the triterpenic molecule, triene **11** was submitted to oxidation processes. This product was obtained by chemical isomerization with iodine of triene **7** and had the central double bond in *E*-disposition.^{11,12} In this sense, triene **11** was ozonized in



Scheme 5. Reagents and conditions: (a) $O_3/CH_2Cl_2/-78$ °C/10 min/Me₂S/3 h 31 (60%); (b) $O_3/CH_2Cl_2/0$ °C/35 min/Me₂S/1 h 39 (45%); (c) $O_3/CH_2Cl_2/Py/-78$ °C/1 h/Me₂S/3 h 40 (80%); (d) $O_3/CH_2Cl_2/Py/room$ temperature/3 h/Me₂S/5 min 39 (10%) and 41 (10%).

was 38, whereas at room temperature the proportion of both compounds was equal. These results, along with the absence of deacetylated derivative 29 in the ozonolysis of 26 at -40 °C, suggested that lactonization minimized at low temperature. When ozonolysis of 34 was maintained for 6 h at room temperature, the yield of compounds 37 and 38 decreased considerably due to the formation of complex mixtures of very polar products.

According to above-described oxidation results, the double bond between C-8 and C-9 was the most reactive, epoxidizing preferentially from the α face. At higher temperatures, the opposite configuration could be achieved. The same could be said for the C-13/C-14 double bond, since it was epoxidized mainly from the α face. The oxirane group in this position gave rise more easily to hydroxylation and lactonization reactions. Finally, the double bond between C-11 and C-12 was not oxidized, preventing the cleavage of C-ring by this bond.

Table 3. Yields (%) of the ozonolysis of product 34 at different times (h) and temperatures (°C)

Temperature	Time	37	38	
-78	0.2	6	55	
Room temperature	0.5	30	30	
Room temperature	6	15	15	

 CH_2Cl_2 at low temperature (-78 °C), providing two diepoxides, **42** and **43**, in a low yield as well as complex mixtures of fragmented and oxidized compounds, which were difficult to separate chromatographically (Scheme 6).

Compound 42 was epoxidized in C-8/C-9 from the β face, whereas the oxirane group at C-13/C-14 had the opposite configuration. In turn, product 43 was produced by the epoxidation from the α face of the most substituted double bonds of the starting material. Thus, the treatment of triene 11 with MCPBA yielded compounds 42 and 43, the latter being the major product of this reaction, together with product 44. This compound was a diepoxide with opposite configurations in the oxirane groups to those in compound 42.

Finally, the oxidation of triene **11** with NaIO₄/RuCl₃ in acetone/H₂O (5:1) led to the cleavage of the triterpene by the C-ring, resulting a mixture of two aldehydic sesquiterpenes (**45** and **46**). One of which (**45**) was a promising 3β-acetoxy-8-drimen-11-al (Scheme 7). This reaction was carried out under different conditions, summarized in Table 4. The best results were achieved when 4 equiv. of NaIO₄ were added to a solution of product **11** and the process was maintained at room temperature for 30 min. The immediate reduction of these unstable sesquiterpene fragments owing to their aldehydic nature gave rise to the corresponding dihydroxylic sesquiterpenes **47**¹⁷ and **48**. Product **47** had structure of 8-drimenol and



Scheme 6. Reagents and conditions: (a) $O_3/CH_2Cl_2/-78$ °C/1 h/Me₂S/3 h 42 (10%) and 43 (10%); (b) MCPBA/CH₂Cl₂/room temperature/30 min 42 (10%), 43 (50%) and 44 (10%).

therefore could be used as an appropriate synthon to produce other remarkable drimane-related compounds. In turn, product **48** would be a suitable intermediate in the synthesis of phenanthrenes and natural tricyclic triterpenes such as Achilleol B and Camelliol C. The reaction of the aldehydic mixture with NaBH₄ provided the partially reduced fragments **49** and **50**. This way, hydroxyl group at C-3 of product **49** was protected as acetate, avoiding the participation in further processes.

Moreover, triene 16, derived from product 8 by a



Scheme 7. Reagents and conditions: (a) $NaIO_4/RuCl_3/acetone/H_2O$ (5:1) 45 and 46 (Table 4); (b) $LiAlH_4/THF/reflux/1$ h 47 and 48 (95%); (c) $NaBH_4/DMF/room$ temperature/1 h 49 and 50 (95%).

Table 4. Yields (%) of products 45 and 46 obtained by oxidation of triene 11 with $NaIO_4$ and $RuCl_3$ at different temperatures (°C) and times (h)

Т	Substrate:reagent	Time	45+46	
-40	1:2	12	30	
-30	1:6	10	10	
-20	1:4	24	15	
0	1:2	1.5	40	
Room temperature	1:4	0.5	50	

photochemical isomerization process,^{11,12} was ozonized under different conditions, but complex mixtures of products resulted. Therefore, intending to minimize undesirable oxidative cleavages, we protected exocyclic triene **16** at the C-13/C-14 double bond by previous epoxidation (Scheme 8). This process led to a major product, **51**, in which the epoxide group had the opposite configuration to the carboxymethyl group at C-28, and product **52**, whose oxirane group had the opposite disposition as that in compound **51**.

Ozonization in CH_2Cl_2 of the epoxydiene **51** at -80 °C for 5 min rendered four fragments (Scheme 9). Two of these had a 14-carbon skeleton (**53** and **54**) and differed only in the configuration of the epoxide group at C-8/C-11, and the others presented a 16-carbon skeleton (**55** and **56**). These four products were achieved as the result of the C-9/C-11 double-bond rupture of the starting triterpene. It was noteworthy that compounds **53** and **54** were adequate



Scheme 8. Reagents and conditions: (a) MCPBA/CH₂Cl₂/-40 °C/12 h 51 (60%) and 52 (15%).



Scheme 9. Reagents and conditions: (a) $O_3/CH_2Cl_2/-80$ °C/5 min/Me₂S/ 36 h 53 (31%), 54 (3%), 55 (10%) and 56 (35%); (b) NaIO₄/RuCl₃/acetone/ H₂O 5:1/room temperature/2 h 55 (30%) and 57 (23%).

ketoepoxy synthons to obtain drimane and ambra oxide related compounds. Thus, product 55 was produced from the D- and E-rings of the triterpenic molecule while hemiacetal 56 was formed from epoxyaldehyde 55 by the attack of a water molecule present in the reaction medium. Ozonization of epoxydiene 51 was tested under different reaction conditions, which affected the yields of the abovementioned products. The aforementioned conditions (O₃, -80 °C, 5 min) were the optimal ones at the moment Moreover, the treatment of epoxydiene 51 with $NaIO_4/$ RuCl₃ led to epoxyaldehyde 55 and diketone 57, recovering 37% of the unreacted starting material. Finally, the ozonolysis of the minor epoxydiene 52 under the same reaction conditions gave rise to ketoepoxides 53 and 54 and epoxyaldehyde 58, which was the isomer of compound 55 in the epoxide group. In this case, hemiacetal 56 was not obtained. Presumably, the epoxide configuration hindered the entry of the necessary water molecule.

3. Conclusion

Several triterpenic compounds with a diene or triene system were used as starting material in different oxidative processes.

Ozonization of the diene system at low temperature led to oxidized products with epoxide, hydroxyl or lactone groups, showing the different accessibility of the two double bonds of these molecules.

The treatment of a *cis*-triene compound and some derivatives with ozone, $NaIO_4/RuCl_3$ and MCPBA gave rise to the same type of oxidized triterpenic products. In all of them the central double bond in Z-disposition was not affected by the oxidative reaction.

In turn, the oxidation of a *trans*-triene with $NaIO_4/RuCl_3$ at room temperature yielded a mixture of two aldehydic sesquiterpenes by the cleavage of the opened C-ring of the substrate.

Moreover, the protection of the C-13/C-14 double bond of an exocyclic triene by epoxidation and subsequent ozonization rendered 16-carbon fragments and *nor*-sesquiterpene compounds with an epoxydecalone structure. Thus, fragments formed from A- and B-rings of the triterpene

could be used as suitable chiral synthon for the semisynthesis of products related to drimane and ambra oxide. Finally, the fragments produced from D- and E-rings of the triterpene molecule could be appropriate intermediates for the production of phenanthrenes and natural tricyclic triterpenes as Achilleol B and Camelliol C.

4. Experimental

4.1. General

Measurements of NMR spectra (300.13 MHz ¹H and 75.47 MHz ¹³C) were made in CDCl₃ (which also provided the lock signal) using BRUKER AM-300 or ARX-400 spectrometers. The assignments of ¹³C chemical shifts were made with the aid of distortionless enhancement by polarization transfer (DEPT) using a flip angle of 135°. Bruker's programs were used for COSY (45°) and C/H and C/C correlation. IR spectra were recorded on a MATTSON SATELLITE FTIR spectrometer. High-resolution mass spectra were made in a MICROMASS AUTOSPEC-Q spectrometer (EBE geometry). Mps were determined using a Kofler (Reichter) apparatus and are uncorrected. Optical rotations were measured on a Perkin-Elmer 241 polarimeter at 25 °C. All reaction solvents were dried and distilled immediately prior to use; chromatography solvents were distilled prior to use. Commercially available reagents were used without further purification. Silica gel Scharlau 60 $(40-60 \ \mu m)$ was used for flash chromatography. CH₂Cl₂ or CHCl₃ containing increasing amounts of Me₂CO were used as eluents. Analytical plates (silica gel, Merck 60 G) were rendered visible by spraying with H₂SO₄-AcOH, followed by heating to 120 °C.

4.2. Isolation of starting material

Oleanolic and maslinic acids^{2,4} were isolated from olivepressing residues, which were extracted in a Soxhlet with hexane and EtOAc successively. Both products were purified from these mixtures by column chromatography over silica gel and transformed into the corresponding methyl esters with ethereal CH₂N₂ or NaOH-MeI and thus, methyl 3 β -hydroxy-12-oleanen-28-oate⁹ and methyl 2 α ,3 β dihydroxy-12-oleanen-28-oate9 were obtained. Acetylation of these esters with Ac_2O/Py at reflux provided the acetylated derivatives.⁹ The treatment of the methyl esters independently with NBS/AIBN for 30 min at reflux gave rise to dienes 1-4,¹² which were irradiated for 20 min in a borosilicate flask using a 125 W high-pressure Hg street lamp and yielding trienes 5-8.¹² Isomerization with iodine of these trienes led to products $9-12^{12}$ and a photochemical reaction of compounds 5-8 with a 125 W high-pressure Hg street lamp in a quartz flask rendered trienes 13-16.¹²

4.2.1. Ozonolysis of 3. Product **3** (200 mg, 0.4 mmol) was dissolved in 10 mL of CH₂Cl₂, stirred at -78 °C and passed through an O₃ flow of 0.1 L/min (10% O₂-90% O₃). After 15 min, 1.5 mL of Me₂S were added. The mixture was maintained with stirring while being cooled down for 3 h. Then it was evaporated and purified over silica gel, yielding 61 mg (30%) of **17**: white solid; mp 218-220 °C; $[\alpha]_D^{25}=-16$ (*c* 0.7, CHCl₃); IR (CHCl₃): ν 3401, 2933,

1736, 1244 cm⁻¹; ¹H NMR (CDCl₃): δ 5.80 (1H, d, J=7.0 Hz, H-11), 4.44 (1H, dd, $J_1=5.2$ Hz, $J_2=10.4$ Hz, H-3), 3.45 (1H, d, J=7.0 Hz, H-12), 2.65 (1H, dd, J₁=3.1 Hz, J₂=13.6 Hz, H-18), 2.05 (3H, s, COCH₃), 1.32 (3H, s, Me), 1.21 (3H, s, Me), 0.97 (3H, s, Me), 0.93 (3H, s, Me), 0.91 (3H, s, Me), 0.90 (3H, s, Me), 0.83 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.2 (Me), 19.1 (C-6), 19.1 (Me), 20.7 (C-15), 21.4 (COCH₃), 21.9 (C-16), 22.3 (Me), 23.7 (Me), 24.3 (C-2), 24.6 (Me), 26.6 (C-22), 27.9 (Me), 31.0 (C-20), 33.3 (C-7), 33.6 (C-29), 34.5 (C-18), 35.2 (C-21), 36.4 (C-1), 38.0 (C-19), 38.6 and 40.3 (C-4 and C-10), 40.9 (C-8), 41.9 (C-14), 43.4 (C-17), 53.7 (C-5), 72.7 (C-12), 80.3 (C-3), 90.8 (C-13), 118.5 (C-11), 151.9 (C-9), 171.0 (COCH₃), 178.8 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 535.3385 (C₃₂H₄₈O₅Na, calcd 535.3399); 45 mg (20%) of **18**: white solid; mp 117–119 °C; $[\alpha]_D^{25}=31$ (*c* 1, CHCl₃); IR (CHCl₃): ν 2951, 1729, 1251 cm⁻¹; ¹H NMR (CDCl₃): δ 4.41 (1H, dd, $J_1=5.5$ Hz, $J_2=9.2$ Hz, H-3), 3.65 (3H, s, COOCH₃), 3.61 (1H, d, J=3.3 Hz, H-11 or H-12), 2.96 (1H, d, J=3.3 Hz, H-11 or H-12), 2.02 (3H, s, COCH₃), 1.17 (3H, s, 3H-27), 1.08 (3H, s, 3H-25), 1.08 (3H, s, 3H-26), 0.93 (3H, s, 3H-24), 0.91 (3H, s, 3H-29), 0.87 (3H, s, 3H-30), 0.76 (3H, s, 3H-23); ¹³C NMR (CDCl₃): δ 16.5 (C-24), 18.5 (C-26), 20.7 (C-6), 21.3 (C-27), 21.3 (COCH₃), 23.5 (C-16), 23.5 (C-30), 24.3 (C-2), 26.3 (C-15), 27.1 (C-25), 27.3 (C-23), 30.7, 32.5 and 32.7 (C-7, C-21 and C-22), 30.8 (C-20), 32.8 (C-29), 34.2 (C-1), 38.7 (C-10), 39.0 (C-4), 39.8 (C-8), 41.1 (C-19), 42.0 (C-14), 43.7 (C-18), 46.2 (C-17), 46.7 (C-5), 51.8 (COOCH₃), 53.2 and 55.7 (C-11 and C-12), 65.1 (C-13), 65.6 (C-9), 80.9 (C-3), 170.9 (COCH₃), 177.6 (C-28); HRLSIMS, *m*/*z*: [M+Na]⁺ 565.3506 (C₃₃H₅₀O₆Na, calcd 565.3505); 32 mg (15%) of **19**: white solid; mp 220–222 °C; $[\alpha]_D^{25} = -21$ (*c* 1, CHCl₃); IR (CHCl₃): ν 3421, 2946, 1726, 1247 cm⁻¹; ¹H NMR (CDCl₃): δ 5.55 (1H, dd, J_1 =3.7 Hz, J_2 =8.2 Hz, H-15), 4.45 (1H, dd, J_1 =6.0 Hz, J_2 =10.4 Hz, H-3), 3.91 (1H, d, J=4.6 Hz, H-12), 3.57 (3H, s, COOCH₃), 3.39 (1H, d, J=4.6 Hz, H-11), 3.07 (1H, dd, $J_1=3.9$ Hz, $J_2=13.7$ Hz, H-18), 2.39 (1H, dd, J₁=8.2 Hz, J₂=15.0 Hz, H-16), 2.02 (3H, s, COCH₃), 1.30 (3H, s, Me), 1.19 (3H, s, Me), 1.07 (3H, s, Me), 0.94 (3H, s, Me), 0.93 (3H, s, Me), 0.85 (3H, s, Me), 0.85 (3H, s, Me); 13 C NMR (CDCl₃): δ 16.3 (Me), 18.8 (C-6), 21.4 (Me), 21.4 (COCH₃), 22.0 (Me), 23.0 (C-2), 27.4 (Me), 29.3 (C-20), 29.5 (Me), 30.0, 31.2 and 32.8 (C-7, C-21 and C-22), 30.1 (Me), 32.9 (C-29), 33.8 and 35.6 (C-1 and C-16), 34.5 (C-18), 38.0 (C-10), 38.1 (C-19), 38.5 (C-4), 41.0 (C-8), 42.0 (C-13), 49.8 (C-17), 49.9 (C-5), 52.0 (COOCH₃), 56.2 (C-11), 68.3 (C-9), 71.5 (C-12), 80.0 (C-3), 120.0 (C-15), 158.2 (C-14), 170.9 (COCH₃), 178.9 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 565.3500 (C₃₃H₅₀O₆Na, calcd 565.3505) and 15 mg (7%) of 20: white solid; mp $126-128 \,^{\circ}C; \, [\alpha]_D^{25}=0 \, (c \, 1, CHCl_3); IR (CHCl_3): \nu 3421,$ 2943, 1735, 1248 cm⁻¹; ¹H NMR (CDCl₃): δ 4.46 (1H, dd, $J_1=5.3$ Hz, $J_2=10.9$ Hz, H-3), 3.94 (1H, d, J=6.3 Hz, H-11), 3.41 (1H, d, J=6.3 Hz, H-12), 2.47 (1H, dd, J₁=5.8 Hz, J₂=11.9 Hz, H-18), 2.04 (3H, s, COCH₃), 1.30 (3H, s, Me), 1.17 (3H, s, Me), 0.96 (3H, s, Me), 0.94 (3H, s, Me), 0.92 (3H, s, Me), 0.88 (3H, s, Me), 0.88 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.7 (Me), 18.2 (C-6), 18.8 (Me), 19.1 (Me), 20.9 (C-15), 21.2 (Me), 21.3 (COCH₃), 21.7 (C-16), 23.6 (C-2), 24.8 (Me), 26.3 (C-22), 28.0 (C-23), 31.1 (C-20), 33.2 (C-7), 33.5 (C-29), 34.1 (C-21), 34.3 (C-18), 34.7 (C-1), 36.4 (C-19), 38.4 and 39.7 (C-4 and C-10), 40.0

(C-8), 41.8 (C-14), 41.8 (C-17), 51.8 (C-5), 60.8 (C-11), 71.8 (C-12), 72.4 (C-9), 80.2 (C-3), 88.6 (C-13), 171.1 ($COCH_3$), 178.2 (C-28); HRLSIMS, m/z: [M+Na]⁺ 551.3351 ($C_{32}H_{48}O_6$ Na, calcd 551.3348).

4.2.2. Ozonolysis of 4. 250 mg (0.4 mmol) of product 4 were dissolved in 10 mL of MeOH/CH2Cl2 4:1 and stirred at -78 °C. Then an O₃ flow of 0.1 L/min (10% O₂-90% O₃) was passed through the solution for 10 min. After that, 90 mg (0.5 mmol) of thiodipropionic acid were added. The mixture was maintained with stirring for 10 min, evaporated to dryness and chromatographed over silica gel to give 101 mg (40%) of **21**: white solid; mp 240-242 °C; $[\alpha]_{D}^{25} = -47$ (c 1, CHCl₃); IR (CHCl₃): v 3441, 2939, 1742, 1252 cm⁻¹; ¹H NMR (CDCl₃): δ 5.73 (1H, d, J=7.0 Hz, H-11), 5.18 (1H, ddd, $J_1=3.8$ Hz, $J_2=J_3=$ 11.0 Hz, H-2), 4.67 (1H, d, J=11.0 Hz, H-3), 3.41 (1H, d, J=7.0 Hz, H-12), 2.61 (1H, dd, $J_1=3.6$ Hz, $J_2=13.1$ Hz, H-18), 2.03 (3H, s, COCH₃), 1.98 (3H, s, COCH₃), 1.30 (3H, s, Me), 1.29 (3H, s, Me), 0.94 (3H, s, Me), 0.92 (3H, s, Me), 0.90 (3H, s, Me), 0.88 (3H, s, Me), 0.83 (3H, s, Me); ¹³C NMR (CDCl₃): δ 17.1 (Me), 19.0 (C-6), 19.0 (Me), 20.6 (C-15), 20.9 (COCH₃), 21.1 (COCH₃), 21.8 (C-16), 23.2 (Me), 23.6 (Me), 24.5 (Me), 26.5 (C-22), 28.1 (Me), 31.0 (C-20), 33.2 (C-7), 33.5 (C-29), 34.4 (C-18), 34.9 (C-21), 36.3 (C-19), 39.7 (C-10), 40.2 (C-4), 41.8 (C-8), 41.9 (C-14), 43.3 (C-17), 43.8 (C-1), 53.4 (C-5), 70.6 (C-2), 72.5 (C-12), 80.0 (C-3), 90.5 (C-13), 118.6 (C-11), 150.7 (C-9), 170.4 (COCH₃), 170.6 (COCH₃), 178.6 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 593.3460 (C₃₄H₅₀O₇Na, calcd 593.3454); 52 mg (20%) of **22**: white solid; mp 137–139 °C; $[\alpha]_D^{25}=56$ (c 1, CHCl₃); IR (CHCl₃): v 2928, 1742, 1248 cm⁻¹; ¹H NMR (CDCl₃): δ 5.56 (1H, d, J=4.6 Hz, H-11), 5.12 (1H, ddd, $J_1=4.6$ Hz, $J_2=J_3=11.0$ Hz, H-2), 4.70 (1H, d, J=11.0 Hz, H-3), 3.61 (3H, s, COOCH₃), 2.97 (1H, d, J=4.6 Hz, H-12), 2.31 (1H, dd, $J_1=4.6$ Hz, $J_2=12.4$ Hz, H-1), 2.05 (3H, s, COCH₃), 1.99 (3H, s, COCH₃), 1.25 (3H, s, Me), 1.23 (3H, s, Me), 0.94 (3H, s, Me), 0.92 (3H, s, Me), 0.89 (3H, s, Me), 0.89 (3H, s, Me), 0.89 (3H, s, Me); ¹³C NMR (CDCl₃): δ17.6 (Me), 17.7 (C-6), 21.0 (COCH₃), 21.2 (COCH₃), 22.5 (Me), 23.5 (Me), 23.7 and 25.7 (C-15 and C-16), 26.0 (Me), 28.5 (Me), 29.9 (Me), 30.8 (C-20), 32.6 (C-7), 33.0 (C-29), 34.1 and 35.3 (C-21 and C-22), 38.7, 39.3 and 40.5 (C-4, C-8 and C-10), 41.0 (C-19), 42.5 (C-1), 42.9 (C-18), 45.1 (C-14), 46.1 (C-17), 50.9 (C-5), 51.8 (COOCH₃), 52.8 (C-12), 69.4 (C-13), 70.3 (C-2), 80.2 (C-3), 114.5 (C-11), 160.3 (C-9), 170.6 (COCH₃), 170.9 (COCH₃), 177.9 (C-28); HRLSIMS, *m*/*z*: [M+Na]⁺ 607.3607 (C₃₅H₅₂O₇Na, calcd 607.3611); and 31 mg (12%) of **23**: white solid; mp 167–169 °C; $[\alpha]_D^{25}=18$ (c 1, CHCl₃); IR (CHCl₃): v 2950, 1739, 1249 cm⁻¹; ¹H NMR (CDCl₃): δ 5.74 (1H, s, H-11), 5.16 (1H, ddd, J_1 =4.6 Hz, J₂=J₃=11.2 Hz, H-2), 4.72 (1H, d, J=11.2 Hz, H-3), 3.69 $(3H, s, COOCH_3)$, 2.34 (1H, dd, $J_1=4.6$ Hz, $J_2=12.4$ Hz, H-1), 2.05 (3H, s, COCH₃), 2.00 (3H, s, COCH₃), 1.44 (3H, s, Me), 1.37 (3H, s, Me), 1.06 (3H, s, Me), 1.00 (3H, s, Me), 0.93 (3H, s, Me), 0.91 (3H, s, Me), 0.82 (3H, s, Me); ¹³C NMR (CDCl₃): δ17.5 (C-6), 17.8 (Me), 20.9 (COCH₃), 21.2 (COCH₃), 25.7 (Me), 26.6 (C-16), 27.2 (Me), 28.4 (Me), 28.4 (Me), 28.9 (C-15), 29.8 (C-22), 30.1 (Me), 32.0 (C-20), 33.8 (C-29), 33.9 (C-7), 35.3 (C-18), 36.6 and 37.1 (C-19 and C-21), 39.4 and 40.6 (C-4 and C-10), 42.0 (C-1), 42.4 (C-8), 43.6 (C-14), 48.0 (C-17), 50.5 (C-5), 52.1

(COOCH₃), 53.2 (C-13), 69.7 (C-2), 79.8 (C-3), 123.5 (C-11), 170.5 (COCH₃), 170.8 (COCH₃), 176.2 (C-9), 178.2 (C-28), 201.5 (C-12); HRLSIMS, *m*/*z*: [M+Na]⁺ 607.3615 (C₃₅H₅₂O₇Na, calcd 607.3611).

4.2.3. Ozonolysis of 5. Product 5 (235 mg, 0.5 mmol) was dissolved in 15 mL of CH₂Cl₂/Py 2:1, stirred at -78 °C and passed through an O₃ flow lower than 0.1 L/min (50% O₂-50% O₃). After 30 min, 1.5 mL of Me₂S were added. The mixture was maintained with stirring while being cooled down for 3 h. Then it was evaporated and purified over silica gel, yielding 32 mg (12%) of 24: syrup; $[\alpha]_D^{25}=91$ $(c 1, CHCl_3); IR (CHCl_3): \nu 3511, 2948, 1724, 1252 cm^{-1};$ ¹H NMR (CDCl₃): δ 7.02 (1H, d, J=15.1 Hz, H-12), 6.68 (1H, d, J=15.1 Hz, H-11), 3.69 (3H, s, COOCH₃), 3.32 (1H, dd, $J_1=5.1$ Hz, $J_2=10.2$ Hz, H-3), 2.03 (3H, s, Me), 1.21 (3H, s, Me), 1.08 (3H, s, Me), 1.02 (3H, s, Me), 0.91 (3H, s, Me), 0.89 (3H, s, Me), 0.85 (3H, s, Me); ¹³C NMR (CDCl₃): δ 15.7 (Me), 16.7 (Me), 19.6 (Me), 21.6 and 22.8 (C-6 and C-16), 24.1 (Me), 26.5 (C-7), 28.3 (Me), 29.6 (C-2), 29.8 (C-20), 29.9 (Me), 31.0 (C-15), 32.8 (C-29), 33.9 and 34.0 (C-1 and C-22), 34.3 (C-18), 35.6 (C-21), 40.4 and 44.5 (C-4 and C-10), 45.5 (C-19), 47.6 (C-5), 51.5 (C-17), 52.0 (COOCH₃), 65.0 (C-8), 68.4 (C-9), 78.2 (C-3), 125.6 (C-11), 146.1 (C-12), 177.8 (C-28), 204.0 and 208.6 (C-13 and C-14); HRLSIMS, m/z: $[M+Na]^+$ 539.3344 (C₃₁H₄₈O₆Na, calcd 539.3349).

4.2.4. Oxidation of 5 with NaIO₄/RuCl₃. NaIO₄ (110 mg, 0.5 mmol) and RuCl₃.3H₂O (approximately 5 mg) in water (2 mL) were added to a solution of product 5 (95 mg, 0.2 mmol) in CCl₄ (5 mL) and CH₃CN (5 mL). The reaction mixture was stirred at room temperature for 8 h and then diluted with CH₂Cl₂, washed with water and dried over anhydrous Na₂SO₄. The solvent was evaporated at reduced pressure and the residue was chromatographed to obtain 84 mg (80%) of **25**: white solid; mp 232–234 °C; $[\alpha]_D^{25}=27$ (c 0.4, CHCl₃); IR (CHCl₃): v 3416, 2946, 1722 cm⁻¹; ¹H NMR (CDCl₃): δ 5.70 (1H, d, *J*=6.4 Hz, H-12), 5.52 (1H, d, J=6.4 Hz, H-11), 3.67 (3H, s, COOCH₃), 3.23 (1H, dd, J_1 =4.7 Hz, J_2 =10.4 Hz, H-3), 2.86 (1H, dd, J_1 =4.4 Hz, J₂=13.6 Hz, H-18), 1.29 (3H, s, Me), 1.21 (3H, s, Me), 1.01 (3H, s, Me), 1.00 (3H, s, Me), 0.97 (3H, s, Me), 0.92 (3H, s, Me), 0.79 (3H, s, Me); ¹³C NMR (CDCl₃): δ 15.6 (Me), 19.6 (C-6), 19.8 (Me), 23.1 (C-16), 24.2 (Me), 26.8 (Me), 27.1 (C-2), 28.4 (Me), 29.2 (Me), 30.4 (C-20), 33.0, 34.1, 34.3, 36.0, 38.6 and 39.6 (C-1, C-19, C-21, C-22, C-15 and C-7), 33.5 (C-29), 39.4 and 42.3 (C-4 and C-10), 42.9 (C-18), 46.3 (C-5), 46.9 (C-17), 52.0 (COOCH₃), 74.0 and 74.5 (C-8 and C-14), 78.6 (C-3), 95.6 and 103.1 (C-9 and C-13), 124.5 (C-11), 134.8 (C-12), 178.5 (C-28); HRLSIMS, m/z: $[M+Na]^+$ 541.3503 (C₃₁H₅₀O₆Na, calcd 541.3505).

4.2.5. Epoxidation of 5. Product **5** (236 mg, 0.5 mmol) was dissolved in 10 mL of CH₂Cl₂ and 130 mg (0.75 mmol) of MCPBA were added. The resulting mixture was stirred at different temperatures (Table 1). When the reaction finished, the mixture was diluted with CH₂Cl₂, extracted with a solution of FeSO₄, neutralized with NaHCO₃, dried over anhydrous Na₂SO₄ and evaporated to dryness. Depending on the reaction conditions (Table 1), chromatography over silica gel yielded different amounts of **26**, **27**, **28**, **29**, **30** and **31**. Product **26**: syrup; $[\alpha]_D^{25} = 5$ (*c* 1, CHCl₃);

IR (CHCl₃): ν 3482, 2926, 1728 cm⁻¹; ¹H NMR (CDCl₃): δ 6.13 (1H, d, J=13.3 Hz, H-12), 5.60 (1H, d, J=13.3 Hz, H-11), 3.66 (3H, s, COOCH₃), 3.50 (1H, dd, J_1 =3.6 Hz, $J_2=12.7$ Hz, H-18), 3.19 (1H, dd, $J_1=4.2$ Hz, $J_2=10.7$ Hz, H-3), 1.51 (3H, s, 3H-27), 1.18 (3H, s, 3H-26), 1.07 (3H, s, 3H-25), 0.92 (3H, s, 3H-23), 0.87 and 0.83 (3H each, s, 3H-29 and 3H-30), 0.75 (3H, s, 3H-24); ¹³C NMR (CDCl₃): δ 15.3 (C-24), 17.0 (C-6), 18.2 (C-25), 20.8 (C-27), 22.3 (C-26), 23.6 (C-16), 25.0 (C-30), 27.4 (C-7), 28.4 (C-2), 28.6 (C-23), 29.5 (C-15), 30.7 (C-20), 31.9 (C-22), 32.5 (C-29), 33.2 (C-1), 34.4 (C-21), 36.1 (C-18), 38.4 (C-10), 38.6 (C-4), 41.5 (C-5), 42.8 (C-19), 45.1 (C-17), 51.8 (COOCH₃), 60.9 (C-8), 71.9 (C-9), 78.6 (C-3), 126.2 (C-11), 127.6 (C-14), 132.7 (C-13), 134.7 (C-12), 178.4 (C-28); HRLSIMS, *m*/*z*: [M+Na]⁺ 507.3452 (C₃₁H₄₈O₄Na, calcd 507.3450). Product **27**: syrup; $[\alpha]_D^{25} = 72$ (*c* 1, CHCl₃); IR (CHCl₃): ν 3484, 2948, 1728 cm⁻¹; ¹H NMR (CDCl₃) δ 5.90 (1H, d, J=13.6 Hz, H-12), 5.65 (1H, d, J=13.6 Hz, H-11), 3.66 (3H, s, COOCH₃), 3.23 (1H, dd, J₁=4.4 Hz, J₂=11.2 Hz, H-3), 2.49 (1H, dd, J₁=3.6 Hz, J₂=12.7 Hz, H-18), 1.12 (3H, s, 3H-26), 1.10 (3H, s, 3H-25), 1.10 (3H, s, 3H-27), 0.93 (3H, s, 3H-23), 0.93 (3H, s, 3H-29), 0.85 (3H, s, 3H-30), 0.78 (3H, s, 3H-24); ¹³C NMR (CDCl₃): δ 15.1 (C-24), 16.9 (C-6), 18.6 (C-25), 20.3 (C-26), 20.8 (C-27), 22.5 (C-16), 24.2 (C-30), 27.0 (C-7), 28.3 (C-2), 28.4 (C-23), 29.5 (C-15), 30.0 (C-20), 31.4 (C-22), 33.2 (C-29), 34.0 (C-1), 35.0 (C-21), 35.2 (C-19), 35.9 (C-18), 37.2 (C-10), 38.7 (C-4), 41.1 (C-5), 45.5 (C-17), 51.7 (COOCH₃), 61.4 (C-14), 62.4 (C-8), 65.6 (C-13), 71.7 (C-9), 78.8 (C-3), 125.4 (C-11), 133.3 (C-12), 178.4 (C-28); HRLSIMS, m/z: [M+Na]⁺ 523.3340 (C₃₁H₄₈O₅Na, calcd 523.3399). Product **28**: syrup; $[\alpha]_D^{25} = -7$ (*c* 1, CHCl₃); IR (CHCl₃): ν 3488, 2947, 1715 cm⁻¹; ¹H NMR (CDCl₃): δ 5.84 (1H, d, J=12.5 Hz, H-12), 5.70 (1H, d, J=12.5 Hz, H-11), 3.74 (3H, s, COOCH₃), 3.24 (1H, dd, J_1 =3.9 Hz, J_2 =14.3 Hz, H-18), 3.21 (1H, dd, J_1 =4.3 Hz, J_2 =11.6 Hz, H-3), 1.17 (3H, s, 3H-26), 1.11 (3H, s, 3H-27), 1.06 (3H, s, 3H-25), 0.90 (3H, s, 3H-23), 0.90 (3H, s, 3H-29), 0.87 (3H, s, 3H-30), 0.76 (3H, s, 3H-24); ¹³C NMR (CDCl₃): δ 15.2 (C-24), 17.1 (C-6), 18.5 (C-25), 19.8 (C-16), 21.7 (C-26), 21.7 (C-27), 24.9 (C-30), 25.6 (C-15), 27.4 (C-7), 28.7 (C-23), 28.8 (C-2), 30.5 (C-20), 31.9 (C-22), 33.1 (C-29), 33.5 (C-21), 34.7 (C-1), 36.7 (C-18), 37.7 (C-10), 38.2 (C-19), 38.6 (C-4), 41.5 (C-5), 42.5 (C-17), 51.4 (COOCH₃), 57.4 (C-14), 60.0 (C-8), 66.3 (C-13), 72.3 (C-9), 78.4 (C-3), 129.8 (C-12), 130.8 (C-11), 178.6 (C-28); HRLSIMS, m/z: [M+Na]⁺ 523.3340 (C₃₁H₄₈O₅Na, calcd 523.3399). Product 29: white solid; mp 197-199 °C; $[\alpha]_D^{25} = 42$ (c 1, CHCl₃); IR (CHCl₃): v 3282, 2936, 1746, 1079 cm⁻¹; ¹H NMR (CDCl₃): δ 5.81 (1H, d, J=14.1 Hz, H-12), 5.60 (1H, d, J=14.1 Hz, H-11), 3.21 (1H, dd, J₁=2.4 Hz, J₂=9.7 Hz, H-3), 1.27 (3H, s, Me), 1.25 (3H, s, Me), 1.12 (3H, s, Me), 0.99 (3H, s, Me), 0.96 (3H, s, Me), 0.82 (3H, s, Me), 0.80 (3H, s, Me); 13 C NMR (CDCl₃): δ 15.3 (C-24), 16.6 (C-6), 18.5 (C-25), 20.4 (C-16), 20.6 and 20.9 (C-26 and C-27), 24.3 (C-30), 27.0, 27.1, 27.7 and 28.3 (C-2, C-7, C-15 and C-22), 28.5 (C-23), 30.5 (C-20), 33.5 (C-1), 33.5 (C-21), 33.5 (C-29), 35.4 (C-19), 37.2 (C-10), 38.7 (C-4), 40.6 (C-17), 41.4 and 42.2 (C-5 and C-18), 67.2 (C-8), 74.2 (C-9), 74.8 (C-14), 78.5 (C-3), 84.7 (C-13), 120.8 (C-11), 138.3 (C-12), 179.0 (C-28); HRLSIMS, m/z: $[M+Na]^+$ 509.3243 (C₃₀H₄₆O₅Na, calcd 509.3243). Product **30**: white solid; mp 152–154 °C; $[\alpha]_D^{25}=37$ (c 0.5,

CHCl₃); IR (CHCl₃): v 3476, 2947, 1727 cm⁻¹; ¹H NMR (CDCl₃): δ 5.90 (1H, d, J=15.4 Hz, H-12), 5.78 (1H, d, J=15.4 Hz, H-11), 3.68 (3H, s, COOCH₃), 3.21 (1H, dd, J_1 =4.4 Hz, J_2 =11.1 Hz, H-3), 2.48 (1H, dd, J_1 =4.2 Hz, J₂=13.0 Hz, H-18), 1.14 (3H, s, Me), 1.12 (3H, s, Me), 1.12 (3H, s, Me), 0.94 (3H, s, Me), 0.89 (3H, s, Me), 0.80 (3H, s, Me), 0.78 (3H, s, Me); ¹³C NMR (CDCl₃): δ 15.3 (Me), 17.0 (C-6), 17.9 (Me), 19.9 (Me), 20.8 (Me), 23.1 (C-16), 24.0 (Me), 27.3 (C-7), 28.4 (Me), 28.8 (C-2), 29.7 (C-15), 29.8 (C-20), 31.1, 34.1, 34.5 and 35.4 (C-1, C-19, C-21 and C-22), 32.8 (C-29), 34.8 (C-18), 37.0 (C-10), 38.7 (C-4), 41.9 (C-5), 44.8 (C-17), 51.9 (COOCH₃), 64.3 and 64.4 (C-13 and C-14), 68.6 (C-8), 73.2 (C-9), 78.4 (C-3), 126.4 (C-11), 133.1 (C-12), 178.1 (C-28); HRLSIMS, m/z: $[M+Na]^+$ 523.3397 (C₃₁H₄₈O₅Na, calcd 523.3399). Product **31**: white solid; mp 268–270 °C; $[\alpha]_D^{25} = -19$ (*c* 1, CHCl₃); IR (CHCl₃): ν 3449, 2948, 1713 cm⁻¹; ¹H NMR (CDCl₃): δ 6.09 (1H, d, J=13.7 Hz, H-12), 5.71 (1H, d, J=13.7 Hz, H-11), 3.71 (3H, s, COOCH₃), 3.21 (1H, dd, J_1 =4.6 Hz, J_2 =11.1 Hz, H-3), 2.57 (1H, dd, J_1 =3.6 Hz, J₂=13.7 Hz, H-18), 1.24 (3H, s, 3H-26), 1.11 (3H, s, 3H-25), 1.11 (3H, s, 3H-27), 0.92 (3H, s, 3H-23), 0.86 (3H, s, 3H-29), 0.78 (3H, s, 3H-30), 0.77 (3H, s, 3H-24); ¹³C NMR (CDCl₃): δ 15.3 (C-24), 16.8 (C-6), 18.7 (C-25), 21.7 (C-27), 22.2 (C-16), 23.5 (C-26), 27.3 (C-2), 27.4 (C-30), 28.3 (C-7), 28.6 (C-23), 30.7 (C-20), 33.3 (C-29), 33.4, 33.7, 34.0 and 34.2 (C-15, C-21, C-22 and C-1), 37.5 (C-10), 37.9 (C-19), 38.6 (C-4), 41.2 (C-5), 44.1 (C-17), 45.2 (C-18), 51.5 (COOCH₃), 65.4 (C-8), 73.7 (C-9), 74.2 (C-14), 77.7 (C-13), 78.2 (C-3), 123.0 (C-11), 137.2 (C-12), 179.8 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 541.3509 (C₃₁H₅₀O₆Na, calcd 541.3505).

4.2.6. Epoxidation of 26. 240 mg (0.5 mmol) of product 26 were dissolved in 10 mL of CH₂Cl₂ and 130 mg (0.75 mmol) of MCPBA were added. The resulting mixture was stirred at different temperatures (-40 °C and room temperature). When the reaction finished, the mixture was diluted with CH₂Cl₂, extracted with a solution of FeSO₄, neutralized with NaHCO3, dried over anhydrous Na2SO4 and evaporated to dryness. Depending on the reaction conditions (Table 2), chromatography over silica gel yielded different amounts of 27, 28, 29, 31, 32 and 33. Product 32: syrup; $[\alpha]_{D}^{25} = 78$ (c 1, CHCl₃); IR (CHCl₃): ν 3536, 2927, 1728, 1285 cm⁻¹; ¹H NMR (CDCl₃): δ 6.13 (1H, d, J=6.3 Hz, H-12), 5.89 (1H, d, J=6.3 Hz, H-11), 5.57 (1H, dd, J₁=3.5 Hz, J₂=5.0 Hz, H-7), 3.50 (3H, s, COOCH₃), 3.17 (1H, dd, J_1 =5.5 Hz, J_2 =10.3 Hz, H-3), 2.67 (1H, dd, J_1 =3.7 Hz, J_2 =14.5 Hz, H-18), 2.25 (1H, ddd, J_1 =3.5 Hz, J₂=13.6 Hz, J₃=17.6 Hz, H-6), 1.67 (3H, s, Me), 1.15 (3H, s, Me), 0.95 (3H, s, Me), 0.88 (3H, s, Me), 0.86 (3H, s, Me), 0.84 (3H, s, Me), 0.84 (3H, s, Me); ¹³C NMR (CDCl₃): δ 14.9 (Me), 18.8 (Me), 22.6 (C-16), 23.3 (Me), 23.7 (Me), 24.8 (C-6), 27.2 (C-2), 27.3 (Me), 30.8 (C-20), 31.3 (Me), 31.5 (C-15), 33.4 (C-29), 34.1 and 34.2 (C-21 and C-22), 34.9 (C-1), 38.4 (C-19), 39.1 (C-4), 40.5 (C-18), 40.7 (C-10), 44.2 (C-5), 45.0 (C-17), 51.6 (COOCH₃), 74.8 (C-14), 79.1 (C-3), 93.8 (C-13), 96.8 (C-9), 126.9 (C-7), 131.1 (C-11), 132.9 (C-12), 133.8 (C-8), 179.0 (C-28); HRLSIMS, m/z: [M+Na]⁺ 523.3397 (C₃₁H₄₈O₅Na, calcd 523.3399). Product **33**: syrup; $[\alpha]_D^{25}=6$ (*c* 0.7, CHCl₃); IR (CHCl₃): ν 3471, 2945, 1724 cm⁻¹; ¹H NMR (CDCl₃): δ 6.14 (1H, d, J=6.4 Hz, H-12), 5.96 (1H, d, J=6.4 Hz,

H-11), 4.83 (1H, d, J=2.0 Hz, H-26a), 4.55 (1H, d, J=2.0 Hz, H-26b), 3.66 (3H, s, COO*CH*₃), 3.17 (1H, dd, $J_1=4.5$ Hz, $J_2=11.2$ Hz, H-3), 2.67 (1H, dd, $J_1=4.3$ Hz, $J_2=13.9$ Hz, H-18), 1.16 (3H, s, Me), 0.98 (3H, s, Me), 0.89 (3H, s, Me), 0.84 (3H, s, Me), 0.78 (3H, s, Me), 0.76 (3H, s, Me); ¹³C NMR (CDCl₃): δ 15.8 (Me), 18.7 (Me), 22.5 and 23.0 (C-6 and C-16), 23.7 (Me), 27.4 (C-2), 28.5 (Me), 30.6 (C-20), 30.9 (Me), 32.6 (C-15), 32.6 (C-7), 33.0 (C-29), 34.0, 35.1 and 35.2 (C-1, C-21 and C-22), 38.9 (C-19), 39.1 and 42.2 (C-4 and C-10), 44.3 (C-17), 44.3 and 45.2 (C-5 and C-18), 51.8 (COO*CH*₃), 74.6 (C-14), 79.1 (C-3), 95.2 (C-13), 98.3 (C-9), 110.8 (C-26), 130.6 (C-11), 133.3 (C-12), 152.7 (C-8), 179.6 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 523.3395 (C₃₁H₄₈O₅Na, calcd 523.3399).

4.2.7. Acetylation of 26. Product **26** (100 mg, 0.2 mmol) was dissolved in 4 mL of pyridine and 2 mL of Ac₂O and stirred for 1 h at reflux. The reaction mixture was diluted with water, extracted with CH₂Cl₂, washed with saturated aqueous KHSO₄ solution and dried with anhydrous Na₂SO₄. The solvent was evaporated at reduced pressure and the residue was chromatographed on a silica gel column to give 103 mg (95%) of **34**: white solid; mp 99–101 °C; $[\alpha]_D^{25}=65$ (c 0.8, CHCl₃); IR (CHCl₃): v 2949, 1732, 1246 cm⁻¹; ¹H NMR (CDCl₃): δ 6.14 (1H, d, J=13.3 Hz, H-12), 5.58 (1H, d, *J*=13.3 Hz, H-11), 4.47 (1H, dd, *J*₁=4.4 Hz, *J*₂=10.6 Hz, H-3), 3.72 (3H, s, $COOCH_3$), 3.55 (1H, dd, $J_1=3.9$ Hz, J₂=12.7 Hz, H-18), 2.01 (3H, s, COCH₃), 1.50 (3H, s, Me), 1.18 (3H, s, Me), 1.09 (3H, s, Me), 0.88 (3H, s, Me), 0.83 (3H, s, Me), 0.83 (3H, s, Me), 0.81 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.4 (C-24), 16.9 (C-6), 18.3 (C-25), 20.8 (C-27), 21.4 (C-26), 21.4 (COCH₃), 22.3 (C-30), 23.8 (C-16), 23.8 (C-2), 28.2 (C-7), 28.6 (C-23), 29.6 (C-15), 30.7 (C-20), 32.0 (C-22), 32.5 (C-29), 32.8 (C-1), 34.5 (C-21), 35.9 (C-18), 37.5 (C-10), 38.3 (C-4), 41.6 (C-5), 42.8 (C-19), 45.2 (C-17), 52.0 (COOCH₃), 61.0 (C-8), 71.6 (C-9), 80.6 (C-3), 126.0 (C-11), 128.0 (C-14), 132.8 (C-13), 134.9 (C-12), 170.9 (COCH₃), 178.4 (C-28); HRLSIMS, *m*/*z*: [M+Na]⁺ 549.3559 (C₃₃H₅₀O₅Na, calcd 549.3556).

4.2.8. Epoxidation of 34. Product 34 (160 mg, 0.3 mmol) was dissolved in 10 mL of CH₂Cl₂ and 78 mg (0.45 mmol) of MCPBA were added. The resulting mixture was stirred at -20 °C for 3.5 h. When the reaction finished, the mixture was diluted with CH₂Cl₂, extracted with a solution of FeSO₄, neutralized with NaHCO₃, dried over anhydrous Na₂SO₄, evaporated to dryness and purified over silica gel, yielding 66 mg (40%) of **35**: syrup; $[\alpha]_D^{25}=50$ (c 0.5, CHCl₃); IR (CHCl₃): v 2949, 1731, 1246 cm⁻¹; ¹H NMR (CDCl₃): § 5.91 (1H, d, J=13.5 Hz, H-12), 5.66 (1H, d, J=13.5 Hz, H-11), 4.48 (1H, dd, J₁=4.1 Hz, J₂=11.4 Hz, H-3), 3.68 (3H, s, COOCH₃), 2.52 (1H, dd, J_1 =3.0 Hz, J₂=13.1 Hz, H-18), 2.02 (1H, s, COCH₃), 1.13 (3H, s, Me), 1.13 (3H, s, Me), 1.10 (3H, s, Me), 0.98 (3H, s, Me), 0.90 (3H, s, Me), 0.86 (3H, s, Me), 0.82 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.2 (C-24), 16.8 (C-6), 18.7 (C-25), 20.4 (C-26), 20.7 (C-27), 21.3 (COCH₃), 22.6 (C-16), 23.5 (C-7), 24.0 (C-30), 28.1 (C-2), 28.3 (C-23), 29.5 (C-15), 30.0 (C-20), 31.5 (C-22), 33.2 (C-29), 34.2 (C-1), 35.0 (C-21), 35.0 (C-19), 35.9 (C-18), 37.1 and 37.6 (C-4 and C-10), 41.4 (C-5), 45.5 (C-17), 51.7 (COOCH₃), 61.3 (C-14), 62.5 (C-8), 65.6 (C-13), 71.7 (C-9), 80.7 (C-3), 125.2 (C-11), 133.4 (C-12), 170.8 (COCH₃), 178.5 (C-28); HRLSIMS,

m/*z*: [M+Na]⁺ 565.3515 (C₃₃H₅₀O₆Na, calcd 565.3505); 17 mg (10%) of **36**: syrup; $[\alpha]_D^{25} = -10$ (c 1, CHCl₃); IR (CHCl₃): ν 2948, 1729, 1247 cm⁻¹; ¹H NMR (CDCl₃): δ 5.82 (1H, d, J=12.5 Hz, H-12), 5.69 (1H, d, J=12.5 Hz, H-11), 4.43 (1H, dd, J_1 =4.4 Hz, J_2 =11.7 Hz, H-3), 3.73 (3H, s, COOCH₃), 3.21 (1H, dd, J₁=4.2 Hz, J₂=13.5 Hz, H-18), 1.99 (1H, s, COCH₃), 1.16 (3H, s, Me), 1.08 (3H, s, Me), 1.07 (3H, s, Me), 0.88 (3H, s, Me), 0.85 (3H, s, Me), 0.81 (3H, s, Me), 0.78 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.4 (C-24), 17.0 (C-6), 18.4 (C-25), 19.8 (C-16), 21.3 (COCH₃), 21.7 (C-26), 21.7 (C-27), 23.8 (C-15), 24.8 (C-30), 25.6 (C-7), 28.7 (C-2), 28.7 (C-23), 30.4 (C-20), 31.9 (C-22), 33.1 (C-29), 33.5 (C-21), 34.4 (C-1), 36.6 (C-18), 37.5 and 37.6 (C-4 and C-10), 38.2 (C-19), 41.7 (C-5), 42.4 (C-17), 51.4 (COOCH₃), 57.3 (C-14), 59.9 (C-8), 66.0 (C-13), 72.0 (C-9), 80.5 (C-3), 129.5 (C-11), 131.1 (C-12), 170.6 (COCH₃), 178.3 (C-28); HRLSIMS, m/z: [M+Na]⁺ 565.3505 (C₃₃H₅₀O₆Na, calcd 565.3505); 16 mg (10%) of **37**: white solid; mp 183–185 °C; $[\alpha]_D^{25}=46$ (*c* 1, CHCl₃); IR (CHCl₃): *v* 3288, 2948, 1737, 1244 cm⁻¹; ¹H NMR (CDCl₃): δ 5.82 (1H, d, J=14.1 Hz, H-12), 5.60 (1H, d, J=14.1 Hz, H-11), 4.47 (1H, dd, $J_1=3.0$ Hz, $J_2=11.3$ Hz, H-3), 2.03 (3H, s, COCH₃), 1.27 (3H, s, Me), 1.24 (3H, s, Me), 1.14 (3H, s, Me), 1.02 (3H, s, Me), 0.86 (3H, s, Me), 0.84 (3H, s, Me), 0.83 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.4 (C-24), 16.5 (C-6), 18.5 (C-25), 20.4 (C-16), 20.7 and 20.9 (C-26 and C-27), 21.3 (COCH₃), 23.4 (C-2), 24.2 (C-30), 27.1, 27.7 and 28.1 (C-7, C-15 and C-22), 28.4 (C-23), 30.5 (C-20), 33.5, 33.5 and 35.1 (C-1, C-19 and C-21), 33.5 (C-29), 37.1 and 37.6 (C-4 and C-10), 40.6 (C-17), 41.6 and 42.2 (C-5 and C-18), 67.1 (C-8), 74.0 (C-9), 74.8 (C-14), 80.1 (C-3), 84.8 (C-13), 120.7 (C-11), 138.5 (C-12), 170.8 (COCH₃), 179.1 (C-28); HRLSIMS, m/z: [M+Na]⁺ 551.3348 (C₃₂H₄₈O₆Na, calcd 551.3349); and 52 mg (30%) of **38**: white solid; mp 198-200 °C; $[\alpha]_{D}^{25}=4$ (c 0.7, CHCl₃); IR (CHCl₃): v 3458, 2948, 1730, 1248 cm⁻¹; ¹H NMR (CDCl₃): δ 6.10 (1H, d, J=13.7 Hz, H-12), 5.70 (1H, d, J=13.7 Hz, H-11), 4.44 (1H, dd, J₁=4.6 Hz, J₂=11.5 Hz, H-3), 3.73 (3H, s, COOCH₃), 2.57 (1H, dd, $J_1=3.7$ Hz, $J_2=13.6$ Hz, H-18), 2.01 (3H, s, COCH₃), 1.24 (3H, s, Me), 1.13 (3H, s, Me), 1.09 (3H, s, Me), 0.86 (3H, s, Me), 0.84 (3H, s, Me), 0.81 (3H, s, Me), 0.78 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.5 (C-24), 16.7 (C-6), 18.6 (C-25), 21.4 (COCH₃), 21.8 (C-27), 22.1 (C-16), 23.5 (C-26), 23.8 (C-2), 27.4 (C-30), 28.1 (C-7), 28.5 (C-23), 30.7 (C-20), 33.4 (C-29), 33.5, 33.8, 33.9 and 34.0 (C-1, C-15, C-21 and C-22), 37.4 (C-10), 37.5 (C-4), 37.9 (C-19), 41.4 (C-5), 44.2 (C-17), 45.2 (C-18), 51.7 (COOCH₃), 65.2 (C-8), 73.4 (C-9), 74.3 (C-14), 77.1 (C-13), 80.4 (C-3), 122.8 (C-11), 137.6 (C-12), 170.8 (COCH₃), 179.7 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 583.3613 (C₃₃H₅₂O₇Na, calcd 583.3611).

4.2.9. Ozonolysis of 26 in CH₂Cl₂. Product **26** (95 mg, 0.2 mmol) was dissolved in 10 mL of CH₂Cl₂, stirred at -78 °C and passed through an O₃ flow of 0.1 L/min (10% O₂-90% O₃). After 10 min, 1 mL of Me₂S was added. The mixture was maintained with stirring while being cooled down for 3 h. Then it was evaporated and purified over silica gel, yielding 34 mg (36%) of **26** and 61 mg (60%) of **31**.

4.2.10. Ozonolysis of 31. 50 mg (0.1 mmol) of product 31 were dissolved in 5 mL of CH_2Cl_2 and stirred at 0 °C. Then

an O_3 flow of 0.1 L/min (10% O_2 -90% O_3) was passed through the solution for 35 min. After that, 0.5 mL of Me₂S were added. The mixture was maintained with stirring while being cooled down for 1 h, evaporated to dryness and chromatographed on a silica gel column, rendering 23 mg (45%) of **39**: white solid; mp 189–191 °C; $[\alpha]_D^{25} = -31$ (*c* 1, CHCl₃); IR (CHCl₃): v 3461, 2949, 1707 cm⁻¹; ¹H NMR (CDCl₃): δ 6.15 (1H, d, J=13.7 Hz, H-12), 5.73 (1H, d, J=13.7 Hz, H-11), 3.69 (3H, s, COOCH₃), 2.57 (1H, dd, $J_1=3.5$ Hz, $J_2=13.5$ Hz, H-18), 1.28 (3H, s, Me), 1.26 (3H, s, Me), 1.09 (3H, s, Me), 1.01 (3H, s, Me), 1.00 (3H, s, Me), 0.86 (3H, s, Me), 0.78 (3H, s, Me); 13 C NMR (CDCl₃): δ 18.0 (C-6), 18.2 (Me), 21.6 (Me), 21.8 (Me), 22.1 (C-16), 23.5 (Me), 26.5 (Me), 27.4 (C-30), 28.3 (C-7), 30.7 (C-20), 33.3 (C-29), 33.4 (C-1), 33.7, 34.0, 34.2 and 34.5 (C-2, C-15, C-21 and C-22), 37.3 (C-10), 37.9 (C-19), 42.6 (C-5), 44.2 (C-17), 45.1 (C-18), 47.0 (C-4), 51.6 (COOCH₃), 65.5 (C-8), 73.0 (C-9), 74.2 (C-14), 77.7 (C-13), 122.6 (C-11), 138.0 (C-12), 179.7 (C-28), 216.2 (C-3); HRLSIMS, m/z: $[M+Na]^+$ 539.3346 (C₃₁H₄₈O₆Na, calcd 539.3349).

4.2.11. Ozonolysis of 26 in CH₂Cl₂/Py. Product 26 (95 mg, 0.2 mmol) was dissolved in 11 mL of CH₂Cl₂/Py 10:1, stirred at -78 °C and passed through an O₃ flow of 0.1 L/min (10% O₂-90% O₃). After 1 h, 1 mL of Me₂S was added. The mixture was maintained with stirring while being cooled down for 3 h. Then it was evaporated and purified over silica gel, yielding 78 mg (80%) of 40: syrup; $[\alpha]_D^{25} = 42 (c 1, \text{CHCl}_3); \text{ IR (CHCl}_3): \nu 2948, 1727 \text{ cm}^{-1}; {}^{1}\text{H}$ NMR (CDCl₃): δ 5.95 (1H, d, J=13.6 Hz, H-12), 5.68 (1H, d, J=13.6 Hz, H-11), 3.67 (3H, s, COOCH₃), 1.27 (3H, s, Me), 1.17 (3H, s, Me), 1.12 (3H, s, Me), 1.02 (3H, s, Me), 1.02 (3H, s, Me), 0.90 (3H, s, Me), 0.83 (3H, s, Me); ¹³C NMR (CDCl₃): δ 18.1 (C-6), 18.2 (Me), 20.2 (Me), 21.0 (Me), 21.5 (Me), 22.4 (C-16), 24.2 (Me), 26.3 (Me), 28.5 (C-7), 29.6 (C-15), 29.9 (C-20), 31.4 (C-22), 33.1 (C-29), 33.9, 34.2, 35.3 and 35.3 (C-1, C-2, C-19 and C-21), 35.9 (C-18), 37.2 (C-10), 42.8 (C-5), 45.5 (C-4), 47.2 (C-17), 51.8 (COOCH₃), 61.9 (C-14), 62.5 (C-8), 65.9 (C-13), 71.2 (C-9), 125.1 (C-11), 134.2 (C-12), 178.4 (C-28), 216.4 (C-3); HRLSIMS, *m/z*: [M+Na]⁺ 521.3235 (C₃₁H₄₆O₅Na, calcd 521.3243).

4.2.12. Ozonolysis of 40. 100 mg (0.2 mmol) of product 40 were dissolved in 11 mL of CH₂Cl₂/Py 10:1 and an O₃ flow of 0.1 L/min (10% O₂-90% O₃) was passed through the solution for 3 h at room temperature. After that, 1 mL of Me₂S was added and the mixture maintained 5 min with stirring, evaporated to dryness and purified over silica gel to give 11 mg (10%) of 39 and 10 mg (10%) of 41: syrup; $[\alpha]_D^{25} = -35 (c \ 0.8, \text{CHCl}_3); \text{IR} (\text{CHCl}_3): \nu 3266, 2948, 1694,$ 1185 cm⁻¹; ¹H NMR (CDCl₃): δ 5.90 (1H, d, J=13.5 Hz, H-12), 5.83 (1H, d, J=13.5 Hz, H-11), 1.24 (3H, s, Me), 1.21 (3H, s, Me), 1.17 (3H, s, Me), 1.03 (3H, s, Me), 0.99 (3H, s, Me), 0.94 (3H, s, Me), 0.80 (3H, s, Me); ¹³C NMR (CDCl₃): δ 18.2 (C-6), 18.3 (Me), 20.0 (C-16), 22.0 (Me), 22.2 (Me), 25.2 (Me), 26.5 (Me), 26.8 and 28.2 (C-7 and C-15), 27.3 (Me), 31.5 (C-20), 33.1 (C-29), 33.9, 34.1, 34.8, 34.9 and 34.9 (C-1, C-2, C-19, C-21 and C-22), 37.4 (C-10), 42.5 (C-18), 44.1 (C-4), 46.4 (C-5), 47.1 (C-17), 60.8 (C-8), 71.5 (C-9), 71.6 (C-14), 87.8 (C-13), 127.3 (C-11), 129.7 (C-12), 179.5 (C-28), 216.9 (C-3); HRLSIMS, m/z: $[M+Na]^+$ 507.3083 (C₃₀H₄₄O₅Na, calcd 507.3086).

4.2.13. Ozonolysis of 34. Product **34** (105 mg, 0.2 mmol) was dissolved in 10 mL of CH_2Cl_2 and passed through an O_3 flow of 0.1 L/min (10% O_2 –90% O_3) at different temperatures. When the reaction was complete, 1 mL of Me_2S was added and the mixture maintained with stirring while being cooled down. Then it was evaporated and chromatographed on a silica gel column to give different amounts of **37** and **38** depending on the reaction conditions (Table 3).

4.2.14. Ozonolysis of 11. Product 11 (100 mg, 0.2 mmol) was dissolved in 10 mL of CH₂Cl₂, stirred at -78 °C and passed through an O_3 flow lower than 0.1 L/min (10% O_2 -90% O₃). After 1 h, 1 mL of Me₂S was added. The mixture was maintained with stirring while being cooled down for 3 h. Then it was evaporated and purified over silica gel, yielding 11 mg (10%) of **42**: syrup; $[\alpha]_D^{25} = 104$ (c 0.5, CHCl₃); IR (CHCl₃): v 2949, 1647, 1246 cm⁻¹; ¹H NMR (CDCl₃): δ 5.94 (1H, d, J=15.5 Hz, H-12), 5.78 (1H, d, J=15.5 Hz, H-11), 4.49 (1H, dd, $J_1=5.3$ Hz, $J_2=10.6$ Hz, H-3), 3.68 (3H, s, COOCH₃), 2.54 (1H, dd, J₁=4.7 Hz, J₂=12.8 Hz, H-18), 2.04 (3H, s, COCH₃), 1.13 (3H, s, Me), 1.10 (3H, s, Me), 1.01 (3H, s, Me), 0.90 (3H, s, Me), 0.84 (3H, s, Me), 0.83 (3H, s, Me), 0.82 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.1 (Me), 16.6 (C-6), 17.0 (Me), 20.1 (Me), 20.3 (Me), 21.3 (COCH₃), 23.2 (C-16), 24.1 (C-30), 24.7 (C-2), 28.0 (C-23), 29.7 (C-7), 29.8 (C-20), 31.2 (C-15), 32.8 (C-23), 34.1 and 34.9 (C-21 and C-22), 35.3 (C-18), 35.9 (C-1), 36.4 (C-19), 38.1 and 38.3 (C-4 and C-10), 44.9 (C-17), 51.9 (COOCH₃), 52.4 (C-5), 64.2 (C-14), 65.6 (C-8), 68.3 (C-13), 72.8 (C-9), 80.3 (C-3), 128.5 (C-11), 133.0 (C-12), 171.0 (COCH₃), 178.0 (C-28); HRLSIMS, m/z: [M+Na]⁺ 565.3510 (C₃₃H₅₀O₆Na, calcd 565.3505); and 10 mg (10%) of **43**: syrup; $[\alpha]_D^{25} = 47$ (*c* 0.4, CHCl₃); IR (CHCl₃): ν 2949, 1732, 1247 cm⁻¹; ¹H NMR (CDCl₃): δ 5.90 (1H, d, J=15.4 Hz, H-12), 5.80 (1H, d, J=15.4 Hz, H-11), 4.45 (1H, dd, J_1 =4.4 Hz, J_2 =11.0 Hz, H-3), 3.68 (3H, s, COOCH₃), 2.47 (1H, dd, J_1 =3.3 Hz, J_2 =13.2 Hz, H-18), 2.02 (3H, s, COCH₃), 1.14 (3H, s, Me), 1.14 (3H, s, Me), 1.10 (3H, s, Me), 0.89 (3H, s, Me), 0.85 (3H, s, Me), 0.83 (3H, s, Me), 0.79 (3H, s, Me); ${}^{13}C$ NMR (CDCl₃): δ 16.4 (Me), 16.9 (C-6), 18.0 (Me), 19.9 (Me), 20.8 (Me), 21.4 (COCH₃), 23.1 (C-16), 23.7 (C-2), 24.0 (C-30), 28.3 (C-23), 28.6 (C-7), 29.7 (C-15), 29.8 (C-20), 31.1 (C-22), 32.8 (C-29), 34.1, 34.3 and 35.4 (C-1, C-19 and C-21), 34.9 (C-18), 36.9 and 37.6 (C-4 and C-10), 42.0 (C-5), 44.8 (C-17), 51.9 (COOCH₃), 64.2 and 64.3 (C-8 and C-14), 68.7 (C-13), 73.0 (C-9), 80.4 (C-3), 126.2 (C-11), 133.3 (C-12), 170.9 (COCH₃), 178.0 (C-28); HRLSIMS, m/z: [M+Na]⁺ 565.3502 (C₃₃H₅₀O₆Na, calcd 565.3505).

4.2.15. Epoxidation of 11. 255 mg (0.5 mmol) of product 11 were dissolved in 10 mL of CH₂Cl₂ and 169 mg (1 mmol) of MCPBA were added. The resulting mixture was stirred at room temperature for 30 min and then diluted with CH₂Cl₂, extracted with a solution of FeSO₄, neutralized with NaHCO₃, dried over anhydrous Na₂SO₄, evaporated at reduced pressure and chromatographed on a silica gel column to obtain 27 mg (10%) of 42, 135 mg (50%) of 43 and 28 mg (10%) of 44: syrup; $[\alpha]_D^{25}=(c \ 1, CHCl_3)$; IR (CHCl₃): ν 2949, 1732, 1246 cm⁻¹; ¹H NMR (CDCl₃): δ 5.94 (1H, d, *J*=3.5 Hz, H-11), 5.94 (1H, d, *J*=3.5 Hz, H-12), 4.49 (1H, dd, *J*₁=4.3 Hz, *J*₂=10.9 Hz, H-3), 3.68 (3H, s, COOCH₃), 2.71 (1H, dd, *J*₁=3.8 Hz,

*J*₂=13.3 Hz, H-18), 2.03 (3H, s, CO*CH*₃), 1.13 (3H, s, Me), 1.09 (3H, s, Me), 1.07 (3H, s, Me), 0.94 (3H, s, Me), 0.91 (3H, s, Me), 0.86 (3H, s, Me), 0.83 (3H, s, Me); ¹³C NMR (CDCl₃): δ 16.2 (Me), 16.7 (C-6), 17.9 (Me), 19.8 (C-16), 21.2 (CO*CH*₃), 21.2 (Me), 21.2 (Me), 23.8 (C-2), 24.6 (C-30), 26.1 (C-15), 28.4 (C-23), 28.7 (C-7), 30.4 (C-20), 32.4 (C-22), 33.0 (C-29), 33.4 (C-21), 34.1 (C-1), 36.9 and 37.8 (C-4 and C-10), 37.6 (C-19), 39.2 (C-18), 42.1 (C-5), 43.0 (C-17), 51.5 (COO*CH*₃), 61.8 (C-14), 64.5 (C-8), 68.5 (C-13), 73.2 (C-9), 80.4 (C-3), 128.0 (C-11), 130.9 (C-12), 170.8 (*CO*CH₃), 178.1 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 565.3495 (C₃₃H₅₀O₆Na, calcd 565.3505).

4.2.16. Oxidation of 11 with NaIO₄/RuCl₃. Product 11 (102 mg, 0.2 mmol) was dissolved in 10 mL of acetone, and a solution in water (2 mL) of approximately 5 mg of RuCl₃.3H₂O and different amounts of NaIO₄ depending on the reaction conditions was added (Table 4). The reaction mixture was stirred at several temperatures and then diluted with CH₂Cl₂, washed with water and dried over anhydrous Na₂SO₄. The solvent was evaporated at reduced pressure and the residue was chromatographed to give variable amounts of 45 and 46 (Table 4). Product 45: white solid; mp 79-81 °C; $[\alpha]_D^{25}=5$ (c 0.4, CHCl₃); IR (CHCl₃): v 2948, 1734, 1245 cm⁻¹; ¹H NMR (CDCl₃): δ 10.03 (1H, s, C-11), 4.50 (1H, dd, J₁=5.5 Hz, J₂=10.9 Hz, H-3), 2.66 (1H, ddd, $J_1 = J_2 = 3.7$ Hz, $J_3 = 13.5$ Hz, H-1 α), 2.04 (3H, s, COCH₃), 2.04 (3H, s, 3H-12), 1.18 (3H, s, 3H-15), 0.89 (3H, s, 3H-13), 0.89 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 16.7 (C-14), 18.0 (C-6), 19.0 (C-12), 20.0 (C-15), 21.3 (COCH₃), 24.1 (C-2), 28.3 (C-13), 34.0 (C-7), 36.7 (C-1), 37.3 (C-10), 37.9 (C-4), 51.2 (C-5), 80.5 (C-3), 143.0 (C-8), 154.6 (C-9), 171.0 (COCH₃), 192.1 (C-11); HRLSIMS, m/z: [M+Na]⁺ 301.1779 (C₁₇H₂₆O₃Na, calcd 301.1780). Product **46**: syrup; $[\alpha]_D^{25} = 51$ (c 1, CHCl₃); IR (CHCl₃): ν 2949, 1727, 1668 cm⁻¹; ¹H NMR (CDCl₃): δ 10.03 (1H, s, C-12), 3.59 (3H, s, COOCH₃), 3.32 (1H, dd, J_1 =2.8 Hz, J_2 =12.2 Hz, H-5), 2.04 (3H, s, 3H-11), 1.03 (3H, s, 3H-13), 0.86 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 18.0 (C-11), 22.2 (C-9), 24.0 (C-13), 30.8 (C-5), 30.9 (C-3), 31.7 (C-8), 32.3 (C-1), 32.8 (C-14), 33.9 (C-2), 42.0 (C-4), 44.8 (C-10), 51.9 (COOCH₃), 137.4 (C-7), 154.6 (C-6), 177.7 (C-15), 190.2 (C-12); HRLSIMS, *m*/*z*: [M+Na]⁺ 287.1617 (C₁₆H₂₄O₃Na, calcd 287.1623).

4.2.17. Obtention of 47 and 48. The oxidation reaction of product 11 with NaIO₄/RuCl₃·3H₂O was worked-up and the residue dissolved in 10 mL of THF without further purification. Then, 2 mL of a solution 1 M of LiAlH₄ in THF were added. The reaction was maintained 1 h at reflux, diluted with aqueous ether, extracted with CH₂Cl₂, dried over anhydrous Na₂SO₄ and evaporated to dryness. Chromatography over silica gel rendered 95% of the reduced products **47**¹⁷ and **48**: syrup; $[\alpha]_{D}^{25}=29$ (c 0.8, CHCl₃); IR (CHCl₃): v 3328, 2916, 1464, 1022 cm⁻¹; ¹H NMR (CDCl₃): δ 4.22 (1H, d, J=11.2 Hz, H-12a), 3.90 (1H, d, J=11.2 Hz, H-12b), 3.42 (1H, d, J=10.8 Hz, H-15a), 3.28 (1H, d, J=10.8 Hz, H-15b), 1.66 (3H, s, 3H-11), 0.91 and 0.89 (3H each, s, 3H-13 and 3H-14); ¹³C NMR (CDCl₃): δ 18.5 (C-11), 22.4 (C-9), 24.2 (C-13), 29.1 (C-1), 30.1 (C-8), 30.9 (C-3), 33.1 (C-14), 34.1 (C-2), 35.3 (C-5), 36.1 (C-10), 42.5 (C-4), 61.6 (C-12), 68.7 (C-15), 130.5 (C-7), 133.1

(C-6); HRLSIMS, m/z: $[M+Na]^+$ 261.1834 (C₁₅H₂₆O₂Na, calcd 261.1830).

4.2.18. Obtention of 49 and 50. Product 11 (102 mg, 0.2 mmol) was dissolved in 10 mL of acetone and a solution in water (2 mL) of approximately 5 mg of RuCl₃·3H₂O and 171 mg (0.8 mmol) of NaIO₄ was added. The reaction mixture was stirred 30 min at room temperature and then diluted with CH₂Cl₂, washed with water, dried over anhydrous Na₂SO₄ and the solvent evaporated to dryness. Without further purification, the residue was dissolved in 5 mL of DMF and 10 mg (0.3 mmol) of NaBH₄ were added. The reaction was maintained 1 h at room temperature and then suspension was evaporated adding 60 mL of toluene/ ethyl ether (3:1). The residue was extracted with CH_2Cl_2 , washed with water, dried over anhydrous Na₂SO₄ and finally evaporated at reduced pressure. Chromatography over silica gel yielded 95% of products 49 and 50. Product **49**: syrup; $[\alpha]_D^{25} = 54$ (*c* 1, CHCl₃); IR (CHCl₃): ν 3447, 2946, 1733, 1246 cm⁻¹; ¹H NMR (CDCl₃): δ 4.48 (1H, dd, J_1 =4.8 Hz, J_2 =11.4 Hz, H-3), 4.16 (1H, d, J=11.6 Hz, H-11a), 4.00 (1H, d, J=11.6 Hz, H-11b), 2.02 (3H, s, COCH₃), 1.89 (1H, ddd, $J_1=J_2=3.5$ Hz, $J_3=12.9$ Hz, H-1a), 1.69 (3H, s, 3H-12), 0.97 (3H, s, 3H-15), 0.86 (3H, s, 3H-13), 0.85 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 16.6 (C-14), 18.6 (C-6), 19.3 (C-12), 20.8 (C-15), 21.4 (COCH₃), 24.1 (C-2), 28.2 (C-13), 33.8 (C-7), 34.6 (C-1), 37.8 (C-10), 37.8 (C-4), 51.0 (C-5), 58.3 (C-11), 80.8 (C-3), 132.7 (C-8), 140.1 (C-9), 171.1 (COCH₃); HRLSIMS, m/z: [M+Na]⁺ 303.1935 (C₁₇H₂₈O₃Na, calcd 303.1936). Product **50**: syrup; $[\alpha]_{D}^{25}=47$ (c 1, CHCl₃); IR (CHCl₃): ν 3463, 2947, 1725, 1253 cm⁻¹; ¹H NMR (CDCl₃): δ 4.17 (1H, d, J=11.1 Hz, H-12a), 3.88 (1H, d, J=11.1 Hz, H-12b), 3.61 $(3H, s, COOCH_3)$, 2.71 (1H, dd, $J_1=3.9$ Hz, $J_2=12.8$ Hz, H-5), 1.60 (3H, s, 3H-11), 0.91 (3H, s, 3H-13), 0.87 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 18.6 (C-11), 22.9 (C-9), 24.3 (C-13), 30.0 (C-8), 30.7 (C-3), 31.7 (C-1), 32.9 (C-14), 34.0 (C-2), 35.6 (C-5), 42.0 (C-4), 45.7 (C-10), 51.9 (COOCH₃), 61.5 (C-12), 130.6 (C-7), 133.8 (C-6), 178.6 (C-15); HRLSIMS, m/z: [M+Na]⁺ 289.1775 (C₁₆H₂₆O₃Na, calcd 289.1780).

4.2.19. Epoxidation of 16. Product 16 (500 mg, 0.9 mmol) was dissolved in 50 mL of CH₂Cl₂ and 228 mg (1.3 mmol) of MCPBA were added. The resulting mixture was stirred at -40 °C for 12 h and then diluted with CH₂Cl₂, extracted with a solution of FeSO₄, neutralized with NaHCO₃, dried over anhydrous Na₂SO₄ and evaporated at reduced pressure to give 310 mg (60%) of **51**: white solid; mp 133–135 °C; $[\alpha]_D^{25} = -36 (c \ 1, \text{CHCl}_3); \text{ IR (CHCl}_3): \nu 3459, 2946, 1743,$ 1368, 1251 cm⁻¹; ¹H NMR (CDCl₃): δ 5.18 (1H, ddd, J_1 =4.6 Hz, J_2 = J_3 =10.3 Hz, H-2), 5.18 (1H, dd, J_1 =4.9 Hz, J₂=9.3 Hz, H-11), 5.02 (1H, d, J=2.2 Hz, H-26a), 4.76 (1H, d, J=10.3 Hz, H-3), 4.59 (1H, d, J=2.2 Hz, H-26b), 3.66 $(3H, s, COOCH_3)$, 2.72 (1H, dd, $J_1=9.3$ Hz, $J_2=15.1$ Hz, H-12a), 2.11 (1H, dd, J_1 =4.9 Hz, J_2 =15.1 Hz, H-12b), 2.04 (3H, s, COCH₃), 2.00 (3H, s, COCH₃), 1.24 (3H, s, Me), 1.07 (3H, s, Me), 0.91 (3H, s, Me), 0.89 (3H, s, Me), 0.86 (3H, s, Me), 0.83 (3H, s, Me); ¹³C NMR (CDCl₃): δ 17.5 (Me), 19.9 (Me), 21.0 (Me), 21.3 (COCH₃), 21.4 (COCH₃), 22.9 (C-6), 24.3 (Me), 28.6 (Me), 29.8 and 30.3 (C-15 and C-16), 29.9 (C-20), 31.6, 34.1 and 33.5 (C-12, C-21 and C-22), 32.8 (Me), 33.7 (C-18), 36.1 (C-19), 36.8 (C-7), 39.7

(C-4), 40.3 (C-1), 41.6 (C-10), 45.1 (C-17), 51.7 and 52.3 (C-5 and COOCH₃), 62.5 and 67.8 (C-13 and C-14), 70.7 (C-2), 80.5 (C-3), 112.7 (C-26), 115.0 (C-11), 144.5 (C-8), 150.7 (C-9), 170.6 (COCH₃), 170.8 (COCH₃), 177.5 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 607.3611 (C₃₅H₅₂O₇Na, calcd 607.3611); and 78 mg (15%) of **52**: syrup; $[\alpha]_D^{25} = -31$ (c 1, CHCl₃); IR (CHCl₃): v 2946, 2857, 1743, 1250, 1035 cm⁻¹; ¹H NMR (CDCl₃): δ 5.22 (1H, dd, J_1 =4.3 Hz, J_2 =10.0 Hz, H-11), 5.19 (1H, ddd, $J_1=3.6$ Hz, $J_2=J_3=10.3$ Hz, H-2), 5.03 (1H, d, J=2.2 Hz, H-26a), 4.75 (1H, d, J=10.3 Hz, H-3), 4.59 (1H, d, J=2.2 Hz, H-26b), 3.67 (3H, s, $COOCH_3$), 2.78 (1H, dd, $J_1=4.3$ Hz, $J_2=14.6$ Hz, H-12a), 2.76 (1H, dd, J₁=10.0 Hz, J₂=14.6 Hz, H-12b), 2.06 (3H, s, COCH₃), 1.99 (3H, s, COCH₃), 1.20 (3H, s, Me), 1.07 (3H, s, Me), 0.93 (3H, s, Me), 0.92 (3H, s, Me), 0.91 (3H, s, Me), 0.88 (3H, s, Me); ¹³C NMR (CDCl₃): δ 17.6 (Me), 19.5 (C-6), 20.7 (Me), 21.0 (COCH₃), 21.2 (COCH₃), 21.6 (Me), 22.8 and 26.5 (C-15 and C-16), 25.2 (Me), 28.8 (Me), 29.8, 32.0 and 33.5 (C-12, C-21 and C-22), 30.6 (C-20), 33.2 (Me), 35.1 (C-18), 36.6 (C-19), 37.2 (C-7), 39.8, 41.5 and 42.9 (C-4, C-10 and C-17), 41.1 (C-1), 51.3 and 52.4 (C-5 and COOCH₃), 61.2 and 68.4 (C-13 and C-14), 70.4 (C-2), 80.6 (C-3), 113.7 (C-26), 114.5 (C-11), 143.8 (C-8), 151.3 (C-9), 170.5 (COCH₃), 170.8 (COCH₃), 178.2 (C-28); HRLSIMS, *m/z*: [M+Na]⁺ 607.3612 (C₃₅H₅₂O₇Na, calcd 607.3611).

4.2.20. Ozonolysis of 51. 250 mg (0.4 mmol) of product 51 were dissolved in 12 mL of CH₂Cl₂ and an O₃ flow of 0.1 L/min (10% O_2 -90% O_3) was passed through the solution for 5 min at -80 °C. After that, 1 mL of Me₂S was added and the mixture maintained 36 h with stirring. Evaporation to dryness and purification over silica gel gave 90 mg (31%) of 53: white solid; mp 154-156 °C; $[\alpha]_{\rm D}^{25} = -12 \ (c \ 1, \ {\rm CHCl}_3); \ {\rm IR} \ ({\rm CHCl}_3): \ \nu \ 2933, \ 1742, \ 1371,$ 1245 cm⁻¹; ¹H NMR (CDCl₃): δ 5.13 (1H, ddd, J_1 =4.5 Hz, J₂=10.3 Hz, J₃=12.1 Hz, H-2), 4.73 (1H, d, J=10.3 Hz, H-3), 3.25 (1H, d, J=5.2 Hz, H-11a), 2.59 (1H, d, J=5.2 Hz, H-11b), 2.18 (1H, dd, J₁=4.5 Hz, J₂=13.5 Hz, H-1β), 2.06 (3H, s, COCH₃), 1.99 (3H, s, COCH₃), 1.36 (3H, s, 3H-12), 1.05 (3H, s, 3H-13), 0.95 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 18.0 (C-14), 18.3 (C-12), 18.6 (C-6), 20.9 (COCH₃), 21.1 (COCH₃), 28.3 (C-13), 31.5 (C-7), 37.3 (C-1), 40.2 (C-4), 49.8 (C-5), 49.9 (C-10), 50.1 (C-11), 57.7 (C-8), 69.0 (C-2), 79.5 (C-3), 170.4 (COCH₃), 170.7 (COCH₃), 207.9 (C-9); HRLSIMS, *m/z*: [M+Na]⁺ 361.1626 (C₁₈H₂₆O₆Na, calcd 361.1627); 10 mg (3%) of **54**: white solid; mp 172–174 °C; $[\alpha]_D^{25} = -104 (c 1, CHCl_3);$ IR (CHCl₃): v 2951, 1740, 1370, 1246 cm⁻¹; ¹H NMR (CDCl₃): δ 5.10 (1H, ddd, J_1 =4.5 Hz, J_2 =10.3 Hz, J₃=12.1 Hz, H-2), 4.73 (1H, d, J=10.3 Hz, H-3), 2.84 (1H, d, J=6.2 Hz, H-11a), 2.73 (1H, d, J=6.2 Hz, H-11b), 2.15 (1H, dd, J_1 =4.5 Hz, J_2 =13.5 Hz, H-1), 2.04 (3H, s, COCH₃), 1.98 (3H, s, COCH₃), 1.28 (3H, s, 3H-12), 1.02 (3H, s, 3H-13), 0.95 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 18.0 (C-13), 18.3 (C-12), 19.9 (C-6), 20.8 (COCH₃), 21.0 (COCH₃), 28.3 (C-14), 32.3 (C-7), 36.9 (C-1), 40.0 (C-4), 49.9 (C-10), 50.0 (C-5), 56.1 (C-11), 58.5 (C-8), 69.0 (C-2), 79.2 (C-3), 170.4 (COCH₃), 170.5 (COCH₃), 207.5 (C-9); HRLSIMS, *m/z*: [M+Na]⁺ 361.1624 (C₁₈H₂₆O₆Na, calcd 361.1627); 25 mg (10%) of **55**: syrup; $[\alpha]_D^{25}=51$ (c 1, CHCl₃); IR (CHCl₃): v 2949, 2865, 1727, 1464, 1254, 1200, 1042 cm⁻¹; ¹H NMR (CDCl₃): δ 9.87 (1H, dd, J_1 =2.7 Hz,

J₂=3.6 Hz, H-13), 3.65 (3H, s, COOCH₃), 2.79 (1H, dd, J_1 =3.6 Hz, J_2 =15.4 Hz, H-12a), 2.44 (1H, dd, J_1 =2.7 Hz, $J_2=15.4$ Hz, H-12b), 2.37 (1H, dd, $J_1=5.8$ Hz, $J_2=11.0$ Hz, H-5), 1.91 (1H, ddd, $J_1=0.0$ Hz, $J_2=4.8$ Hz, $J_3=12.7$ Hz, H-8a), 1.76 (1H, ddd, J_1 =0.0 Hz, J_2 =4.8 Hz, J_3 =12.7 Hz, H-8b), 1.29 (3H, s, 3H-11), 0.92 and 0.86 (3H each, s, 3H-14 and 3H-15); ¹³C NMR (CDCl₃): δ 20.1 (C-11), 22.9 (C-9), 24.0 (C-14), 29.8 (C-3), 29.8 (C-8), 30.9 (C-1), 32.7 (C-15), 33.9 (C-2), 35.4 (C-4), 35.7 (C-5), 45.2 (C-10), 49.6 (C-12), 51.9 (COOCH₃), 62.3 (C-6), 64.1 (C-7), 177.8 (C-16), 200.6 (C-13); HRLSIMS, *m*/*z*: [M+Na]⁺ 317.1732 (C₁₇H₂₆O₄Na, calcd 317.1729); and 95 mg (35%) of **56**: syrup; $[\alpha]_D^{25} = -24$ (c 1, CHCl₃); IR (CHCl₃): v 3450, 2950, 1726, 1257, 1036, 977 cm⁻¹; ¹H NMR (CDCl₃): δ 5.20 (1H, dd, J_1 = J₂=5.8 Hz, H-13), 3.71 (3H, s, COOCH₃), 2.51 (1H, dd, $J_1 = 5.8 \text{ Hz}, J_2 = 14.0 \text{ Hz}, \text{ H-12a}, 2.43 \text{ (1H, dd, } J_1 =$ $J_2 = 8.9$ Hz, H-5), 2.13 (1H, dd, $J_1 = 5.8$ Hz, $J_2 = 14.0$ Hz, H-12b), 1.31 (3H, s, 3H-11), 0.91 (3H, s, 3H-14), 0.91 (3H, s, H-15); ¹³C NMR (CDCl₃): δ 22.0 (C-9), 23.9 (C-14), 26.0 (C-11), 30.7 (C-3), 31.4 (C-1), 33.2 (C-15), 33.4 (C-8), 33.5 (C-2), 38.3 (C-4), 41.8 (C-5), 45.7 (C-12), 47.1 (C-10), 52.5 (COOCH₃), 81.7 (C-7), 82.0 (C-6), 95.9 (C-13), 181.2 (C-16); HRLSIMS, *m/z*: [M+Na]⁺ 335.1828 (C₁₇H₂₈O₅Na, calcd 335.1834).

4.2.21. Oxidation of 51 with NaIO₄/RuCl₃. NaIO₄ (171 mg, 0.8 mmol) and RuCl₃·3H₂O (approximately 5 mg) in water (2 mL) were added to a solution of product 51 (100 mg, 0.2 mmol) in acetone (10 mL). The reaction mixture was stirred at room temperature for 3 h and then diluted with CH₂Cl₂, washed with water and dried over anhydrous Na₂SO₄. The solvent was evaporated at reduced pressure and the residue was chromatographed to obtain 37 mg (37%) of **51**, 30 mg (30%) of **55** and 25 mg (23%) of **57**: syrup; $[\alpha]_{D}^{25} = -11$ (c 1, CHCl₃); IR (CHCl₃): ν 3460, 2937, 2857, 1742, 1678, 1370, 1321, 1052 cm⁻¹; ¹H NMR (CDCl₃): δ 6.11 (1H, dd, J₁=3.8 Hz, J₂=5.6 Hz, H-7), 5.14 (1H, ddd, J_1 =4.6 Hz, J_2 =10.4 Hz, J_3 =12.0 Hz, H-2), 4.74 (1H, d, J=10.4 Hz, H-3), 2.06 (3H, s, COCH₃), 2.00 (3H, s, COCH₃), 1.23 (3H, s, 3H-12), 1.07 (3H, s, 3H-13), 0.93 (3H, s, 3H-14); ¹³C NMR (CDCl₃): δ 18.1 (C-13), 18.3 (C-12), 20.9 (COCH₃), 21.1 (COCH₃), 21.5 (C-6), 27.6 (C-14), 36.9 (C-1), 39.7 (C-4), 44.9 (C-10), 48.6 (C-5), 69.0 (C-2), 79.5 (C-3), 116.5 (C-7), 144.6 (C-8), 170.4 (COCH₃), 170.6 $(COCH_3)$, 199.4 (C-9); HRLSIMS, m/z: $[M+Na]^+$ 347.1467 (C₁₇H₂₄O₆Na, calcd 347.1471).

4.2.22. Ozonolysis of 52. 250 mg (0.4 mmol) of product 52 were dissolved in 12 mL of CH₂Cl₂ and an O₃ flow of 0.1 L/min (10% O₂-90% O₃) was passed through the solution for 5 min at -80 °C. After that, 1 mL of Me₂S was added and the mixture maintained 36 h with stirring. Evaporation to dryness and purification over silica gel yielded 89 mg (31%) of 53, 9 mg (3%) of 54 and 115 mg (46%) of **58**: syrup; $[\alpha]_{D}^{25}=19$ (c 1, CHCl₃); IR (CHCl₃): ν 2949, 2865, 1718, 1463, 1260, 1173, 1045 cm⁻¹; ¹H NMR (CDCl₃): δ 9.76 (1H, dd, J_1 =2.0 Hz, J_2 =2.7 Hz, H-13), 3.67 (3H, s, COOCH₃), 2.76 (1H, dd, J_1 =4.0 Hz, J_2 =13.4 Hz, H-5), 2.72 (1H, dd, J_1 =2.7 Hz, J_2 =16.9 Hz, H-12a), 2.56 (1H, dd, J₁=2.0 Hz, J₂=16.9 Hz, H-12b), 1.21 (3H, s, 3H-11), 0.93 (3H, s, 3H-14), 0.93 (3H, s, 3H-15); ¹³C NMR (CDCl₃): δ 19.0 (C-9), 21.1 (C-11), 24.5 (C-14), 26.1 (C-8), 30.5 (C-3), 32.1 (C-1), 33.0 (C-15), 33.2 (C-2), 37.5

(C-4), 38.4 (C-5), 43.1 (C-10), 46.5 (C-12), 51.6 (COO*CH*₃), 60.2 (C-6), 64.9 (C-7), 178.0 (C-16), 200.1 (C-13); HRLSIMS, m/z: [M+Na]⁺ 317.1729 (C₁₇H₂₆O₄Na, calcd 317.1729).

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